
The vomeronasal organ functions in entropy dissipation, the communication by pheromones for a feedback by the pituitary over brain plasticity and the development of the unconscious

Dr. Alfred Bennun
Full Professor Emeritus of Biochemistry
Federated Departments of Biological Sciences
New Brunswick, Rutgers, The State University of New Jersey

Abstract

A model of human brain evolution should take in account the constitutive separation of two integrated parameters: the one for neuronal synapsis and circuits, from that by a multiple wrapped-around astrocytes and their exchanges. The main function of glial cells is the absorption of the generated, thermal-like breaking of H-bonds, from polymerized water provides the activation energy, coupled for the turnover of structural changes, between hydrophobic and hydrophilic enzyme forms. Thus, increasing rotational and vibrational kinetic activity, on the separated individual H₂O molecules, but maintaining a liquid coherence, during circulation within astrocytes until the lower pressure at the vomeronasal organ (VNO) allows phase conversion to vapor, equivalent to entropy dissipation. The summation of the energy generated by metabolites and H-bond consumption allows the brain thermodynamics to support ratios between metabolite concentrations and the electrogenic action potential in dissipative states, within an open system. Cell-free DNA (cffDNA) from an evolutive disaggregation of the olfactory bulb, are still present as an olfactory epithelium, adapted for sex pheromones and behavioral responses stimulus on adenylate cyclase (AC) for cAMP-dependent unzipping of the DNA. It is a focused process to create neurons differentiated by where the cAMP is inserted, which creates specific neural circuits. Olfactory epithelium may hold the neurons with 7TM receptors for the saliva or pheromones, reaching the oral cavity with their axons crossing the BBB. This allows cffDNA response through cAMP complexing and messenger RNA expression into polypeptide holding a transitory memory during baby rearing, as a psychosomatic molecular carrier of emotional and communication needs of a newborn, which functions to form an unconscious level, which diversifies individual emotional characteristics, approaching a level of animic personalities, which when age allows a conscious level, could offer differentiable roles for social influences that allow the brain self-plasticity such as literacy.

Keywords: VNO, dissipation of entropy, pheromones communication, psychosomatic, hypothalamic-adenylate cyclase, cAMP unzipping cffDNA, postnatal memory, psychosexual development complexes, unconscious, autism, oxytocin, feedback on brain evolution patterns.

Introduction

The pheromone communication level from animals is as an integrative function whereas appears as an evolutionary transition stage in humans. Before the development of an active audio-visual language, the neuronal system of the newborns, allows for the memorization of experience, through the scent-smell-touch system of pheromone binding, by seven-transmembrane (7TM) receptors dendrites of neurons with axons crossing the BBB as sensory inputs, but do not allow the crossing of adrenaline

itself. The characterization of 7TM receptors-AC of rat brain hypothalamus [¹], indicated that cAMP, could be involved in pheromone-communication. The kinetic equilibrium of the hypothalamic-adenylate cyclase (AC) [EC 4.6.1.1] produces cAMP for unzipping cell-free fetal DNA (cffDNA) of memory pathways and their transduction into messenger RNA (mRNA), of non-nuclear and non-inheritable origin, psychosomatic carrier of many emotions like fear, attachment, etc.

The human brain has evolved from a mammal smell-olfactory-memory organ, which allows in the short time a capability for continuous coordination and muscle control. In humans a pheromone-communication level remains for a long nurturing time until could be replaced by a new audio-visual recall memory, at the conscious level. This one contains memory from the previous evolutive stage as vestiges into a subconscious level, which structure psychosomatic complexes.

At the stage of nurturing, the creation of a transitory memory could not be the nuclear DNA of neurons of the neuronal circuits.

Thus, the newborn connects a scent-smell-touch communication functions as a psychic level memory. It is clear that pheromone-like chemical signals play a role in offspring identification and mother recognition and sucking pleasure.

The pheromone-dependent activation of hypothalamic-AC interacts with cffDNA, originated from the never develop olfactory bulb cells, as fragments between 50 and 200 base pairs of DNA, and released to the blood plasma. cffDNA becomes increasingly frequent in circulation with the onset of age [²]. In the maternal blood cffDNA is variable, but on average it is 10% of the fetus, 5-7 gestation weeks, but two days after delivery is no longer detectable in maternal blood [³].

Pheromone and hormone carried by air-saliva functions as a primitive molecular communication of recognition and memory, reflecting a previous evolutionary stage, which as signals to the hypothalamic-amygdala-pituitary-adrenal axis. Also the system responds as a relay site to the oxytocin tongue.

Oxytocin is a neuropeptide released by the posterior pituitary [⁴] into portal circulation increases trust and social bonding. It plays a role in sexual reproduction, and childbirth [⁵], maternal behavior and causes sexual arousal, like kissing between lovers (saliva) or breastfeed between mother and baby (milk). Oxytocin signaling triggers a behavioral response to the presence of prey, predators, and sex pheromones.

The bonding by oxytocin [⁶] and other pheromones appear as smell-touch communication memory between mother and its child, for healthy nurturing and rising. Hence, the origin of complexes could be related to the Freudian psychology of psychosexual development stages: 1. the Oral, 2. the Anal, 3. the Phallic, 4. the Latent, and 5. the Genital. This relates the body region of the libido pleasure to a different erogenous zone of the infant's body. The newborn memory of care by their parents would surface as a latter age as the origin of Oedipus and Electra complexes, etc. Low levels of oxytocin, during nurturing, could become a condition leading to a disease, like autism, or repressed anguish and angry feeling.

Brain evolution has been conditioned and circumscribed by transition stage parameters, adjusted by nurturing to energy (nutritional contents) and emotional exchanges, to develop the psychosomatic complexes, like a child's desire for parental care for the opposite-sex parent time.

The VNO operates as a radiator mechanism, dissipating heat captured by the breaking of H-bonds between molecules of H₂O latter circulating. Astrocytes wrapped around neurons carry H₂O independent of acquiring stable microscopic structure phase until released as a 5% vapor of exhaled air preventing reversal equilibrium and preventing an entropy increase.

This thermodynamics approach allows the evolutive increase in efficiency of the 1.6 kg brain to process a 30% of total body ingested calories, disposing of expended CSF-water (poor H-bonds content) by the astrocytes, maintaining a quasi-liquid state at 36.6 °C.

Mass-action effects by Ca²⁺ inhibit AC and activate calcium permeable AMPARs receptor, during the brain state of sleeping. This state of synchrony contributes circuits in a labile state for memory formation. An abnormal synchrony expression during sleep-wake [¹⁴] has been implicated in drug addiction and memory disorders.

A lower caloric diet (or starvation) lowers [ATP⁴] and [CaATP] both dead end inhibitor activating AC, which raises cAMP as a molecular signal driving adaptive evolution and conferring longevity to man and animals.

A technic of assay of cAMP levels could provide a test to manifest organic dysfunctions. Assay of oxytocin could be a test for diagnosis of autism and psychosomatic dysfunctions, which could result from a detached care of newborns, during the breastfeeding period. Moreover, to create a proper commercial mother's milk substitute, may require the addition of pheromones that may contribute to potentiate a desirable ulterior behavior.

The sensory activity of neurons crossing the BBB conditioning of the unconscious

The scent-touch of the newborn of pheromones communication could be characterized as a molecular level origin of the memory persisting as psychosomatic complexes. The axons from neurons, called cranial nerve zero (CN 0), project into the human olfactory epithelium (not detectable as an organ *per se* as is the case of the animals). Saliva or pheromones bind to neurons dendrites show 7TM receptors in the oral cavity with axons cross BBB, without transferring the adrenaline, to allow AC to stimulate the HTPA axis response.

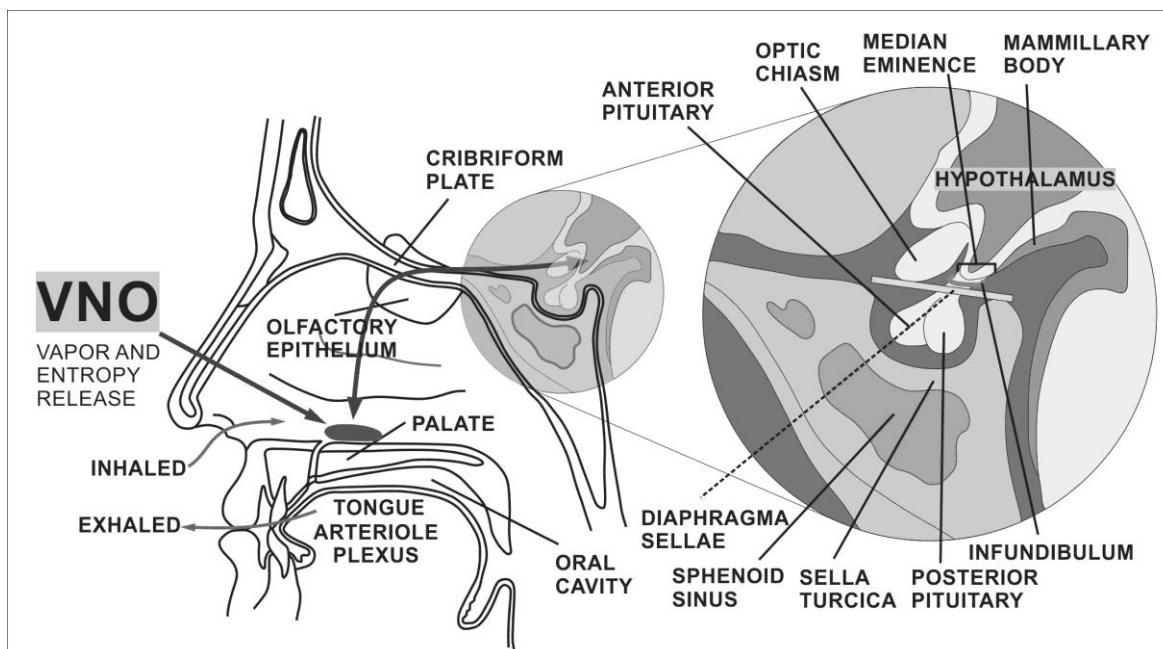


Figure 1: The VNO consists of the paired (two slits), to allows brain to dissipate entropy, which is located in nasal septum at the base of the nasal cavity, located just above the roof of the mouth (the hard palate) (by releasing the CSF depleted of H-bonds that scape from the oral cavity as vapor conforming an exhaust for generated heat of entropy), the epithelium functions as a sponge tissue, exchanging liquids or smells. Hypothalamic-pituitary-adrenal axis (HTPA) shows a bed nucleus of the stria terminalis, serving as a relay site of a major output pathway of the amygdala that responds to threat with anxiety. Oxytocin acting in the mesolimbic dopamine system promotes social approach in positive social contexts. **Hypophyseal fossa in cavity, Sella Turcica:** covered by a flat piece extension of the dura mater (diaphragma sellae), with a circular hole, allowing the vertical passage of the pituitary stalk to the hypothalamus. The latter, controls the energy balance, synthesis, release, activity of anterior pituitary (such as physical and emotional stress, coitus and suckling) and feedbacks by target glands hormones (Thyroxin, Cortisol and Gonadal steroids), through the hypophyseal portal circulation, connecting arterioles (plexus) reach the tongue.

The neurons crossing the BBB are activated by the binding of certain chemicals to their G protein-coupled receptors: they express receptors from three families, called V₁R [7] [8] [9], V₂R, and FPR [10] [11]. The mammalian species encode genes of seven-transmembrane (7TM) G-protein-coupled receptors, which function to identify odorants, pheromones, and saliva-hormones [12], that are characterized by transcription and expression of the sensory neuroepithelium.

The physiological changes of the multiple equilibriums, over AC, allows a signaling predominance of Mg²⁺ over Ca²⁺ or vice versa, connecting with the Ca²⁺ inhibition of AC, allowing the concurrent activation of the α-amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor.

Hypophyseal AC will respond forming the cAMP-Mg-cffDNA complex, at the early stages of a newborn conditioned memory of emotional bonding with their parents.

Hypothalamic-adenylate cyclase activation the HTPA axis secretion connects the portal-small arterioles of the tongue for hormonal release response to incoming pheromones detected by the olfactory epithelium

The isolation from rat's brain AC and characterization for its neurotransmitter responsiveness allowed evaluation of a possible role of the enzyme in cAMP-dependent memory processes. NA activation, when Mg^{2+} is in excess of substrate, of an isolated AC from brain, was first reported for cortex, corpus striatum and hypothalamus [¹³]. The enzyme relates to a network of tissues [¹⁴] supporting brain activity [¹⁵] by their *in common* response to the modification by changes in the concentration of chelating metabolites of free ionic Mg^{2+} (not chelated).

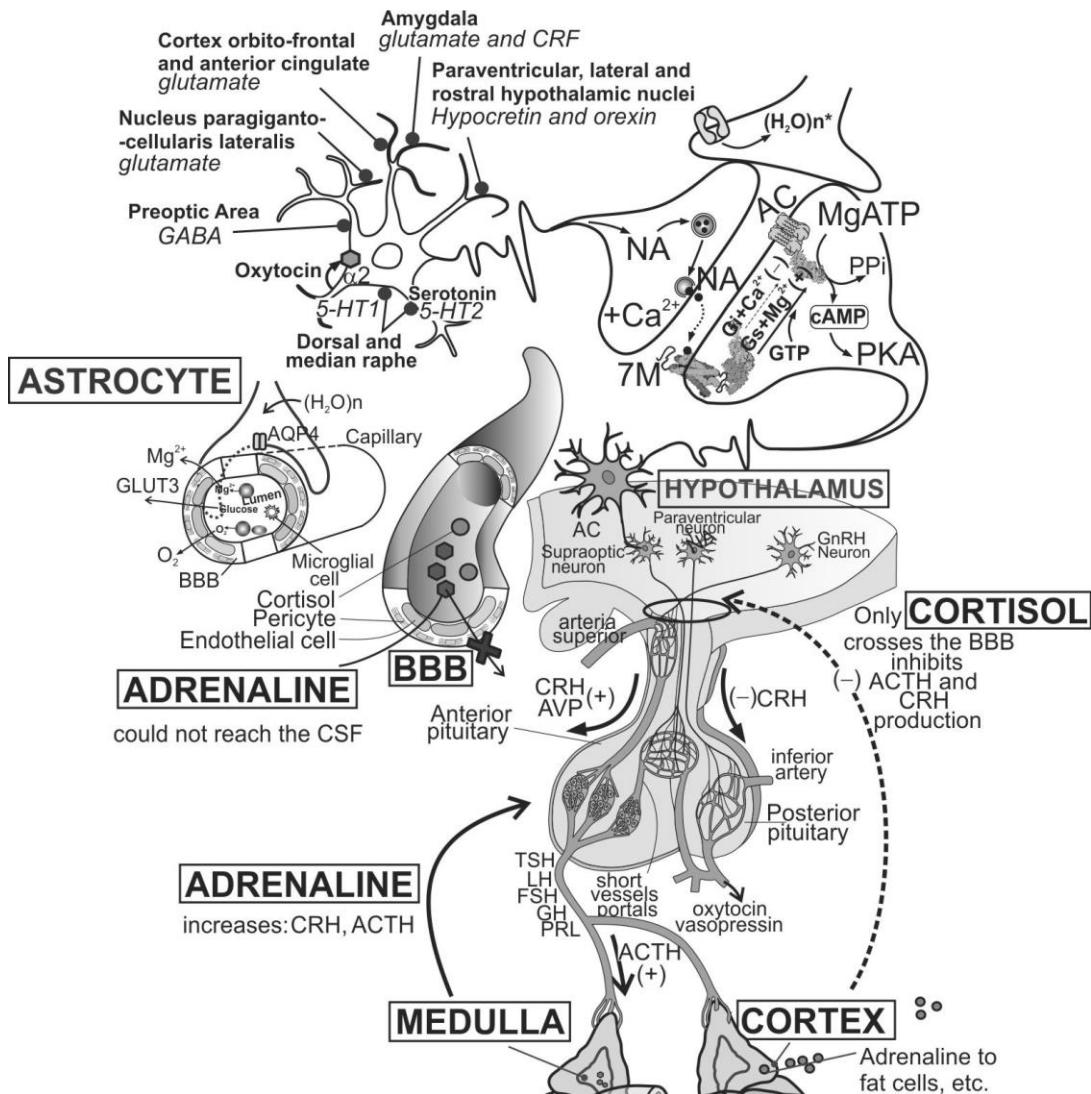


Figure 2: Ionic-metabolic integrative coupling of the HTPA neuronal system. G-protein-coupled receptor (GPCRs) exchange its binding to GDP inactive for GTP active (GDP/GTP-exchange) for NA receptor coupling to AC. Neuron uptake and stoichiometric cycling reversing the glutamate synthase reaction supports release of glutamate (Glu), activating the postsynaptic N-methyl-D-aspartate (NMDA) receptor and ion channel protein. At the astrocyte synaptically released GLU is co-transported with Na^+ by the Na^+/K^+ ATPase consuming one ATP in the exchange for extracellular K^+ . These various effectors are then responsible for AC response of cAMP signaling functions that include control of metabolism, gene transcription and ion channel activity. In many cases, these functions are modulatory in that cAMP often acts to adjust the activity of other signaling pathways and thus has a central role to play in the cross-talk between signaling pathways. This modulatory function is particularly evident in the case of Ca^{2+} signaling in both neuronal and muscle cells to control voltage-gated ion channels finely tuned to regulate many aspects of signal transduction and their dysfunction effect on several disorders [¹⁶] [¹⁷].

Cognition and Memory

The origin of cognition and memory could be attributed to the post-natal scent-smell communication memory of human infants. Degraded cell-free fetal DNA (cffDNA) fragments released to the blood plasma, could not correspond with a characteristic expression of the genetic DNA in the nuclei, which is transmissible to the progeny. The latter, is not transfer by hereditary mechanism, like would be the case of arachnid's spiderweb. This, will involve duplication of DNA preserved in the neuronal nuclei, from where, according to genetics should have been, a marker transmitted by successive generation. Conditioning could on the other hands implicate anticipatory memory, for appetitive which is present as a reflex. The reflex conditioning response described by Pavlov using dog's experimental studies. In animals, the olfactory organ integrates a memory, which allowed their offspring to reach self-care, in a short time.

In the newborn human, the residual structure from evolutional deletion of the olfactory sense allows a memory unable to coordinate muscles. This process requires a long period of parental care, before reaching the brain structure of neuronal circuits, capable to support muscular interaction and development through a cognitive visual-hearing language.

The astrocytes-vomeronasal organ axis function for vapor release

H-bond dissipated water through the astrocytes system eventually could reach the human vomeronasal organ (VNO) [¹⁸] [¹⁹].

The evidence suggests that the increased morphological complexity of astrocytes that has occurred within primate phylogeny may increase cognitive function [²⁰].

The glial cells (astrocytes) [²¹], at the rate of 50:1 per neuron, are the more numerous cells of the neuronal system. A single astrocyte could contact with up to 2 million synapses [²²] [²³]. Astrocytes are wrapped around neurons utilizes the perivascular spaces formed by the vascular endfeet of astrocytes, around the vasculature.

The glymphatic system has a role of maintaining the homeostasis of the central nervous system (CNS), through extensive contact with the cerebral capillaries absorbing heat, during neuronal activatory responses [²⁴] dissipating it outside the system's entropy.

Radial astrocytes can be found between the grey and pia matter and communicate with cerebrospinal fluid (CSF) and help in maintaining the blood-brain barrier (BBB). They release glutamate, D-serine, GABA, and ATP to the neurons. Astrocytes express glutamate transporters, K⁺ channels and water channels, allowing them to contribute to the maintenance of glutamate homeostasis, potassium homeostasis and H-bonds water exchanges homeostasis, respectively [²⁵].

The Hypothalamic-Pituitary-Adrenal Axis Control on the Psychosomatic Metabolic Network

The characterization of the overstimulation of the NA-enzyme AC system (7TM-AC) of the hypothalamic-pituitary-adrenal (HTPA) axis could turn-on the fight-or-flight response. The increment of adrenaline secretion, but without entering the CSF, shifts body metabolism in the direction of depleting metabolic reserves like fats and cortisol releasing amino acids from proteins to support gluconeogenesis. Adrenaline could not cross the blood-brain barrier, to act by negative feedback of its secretion, in the homeostatic metabolic and functional states of brain. Adaptive brain evolution may explain the lack of adrenaline feedback as a restriction to signaling, which allows the dominance of brain over its metabolic supporting tissue network. A molecular perspective could therefore explain the advantage of assigning to the brain unchallenging control, for maximizing the organismal efforts required for survival. This brain pattern of emotional control, over metabolic supporting functions, may participate on the psychosomatic bases of the unconscious.

Pheromone activation of hypothalamic-adenylate cyclase and cAMP unzipping of cffDNA

This conjecture had experimental support from many laboratories and has been successfully generalized as behavior-cAMP linked models [26] [27] [28] [29] [30]. Adrenaline is coupled to the active site in AC that is coupled to 7TM G protein receptors activated by a GTP cycle. Noradrenaline (NA) is released by the long axons of neurons of the locus-coeruleus into the synaptic junctions for sensorial-integrated perception between many brain areas. The activation of the Na⁺/K⁺-ATPase pump release nascent Mg²⁺, by decreasing [ATP⁴⁻], which has an inhibitory effect on AC. The capture by nascent Mg²⁺ of water from the hydration shells of the less strong ions decreases the sizes of Na⁺ and K⁺, fitting both to their gates, allowing across the membrane the sieve effects, which confers specific pattern to the wavelength of the action potential. Brain NA is contained in neuronal junction's vesicles to activate other neurons. NA activated-AC (not to confuse with adrenaline), which is not present in brain are located in the locus-coeruleus system [6].

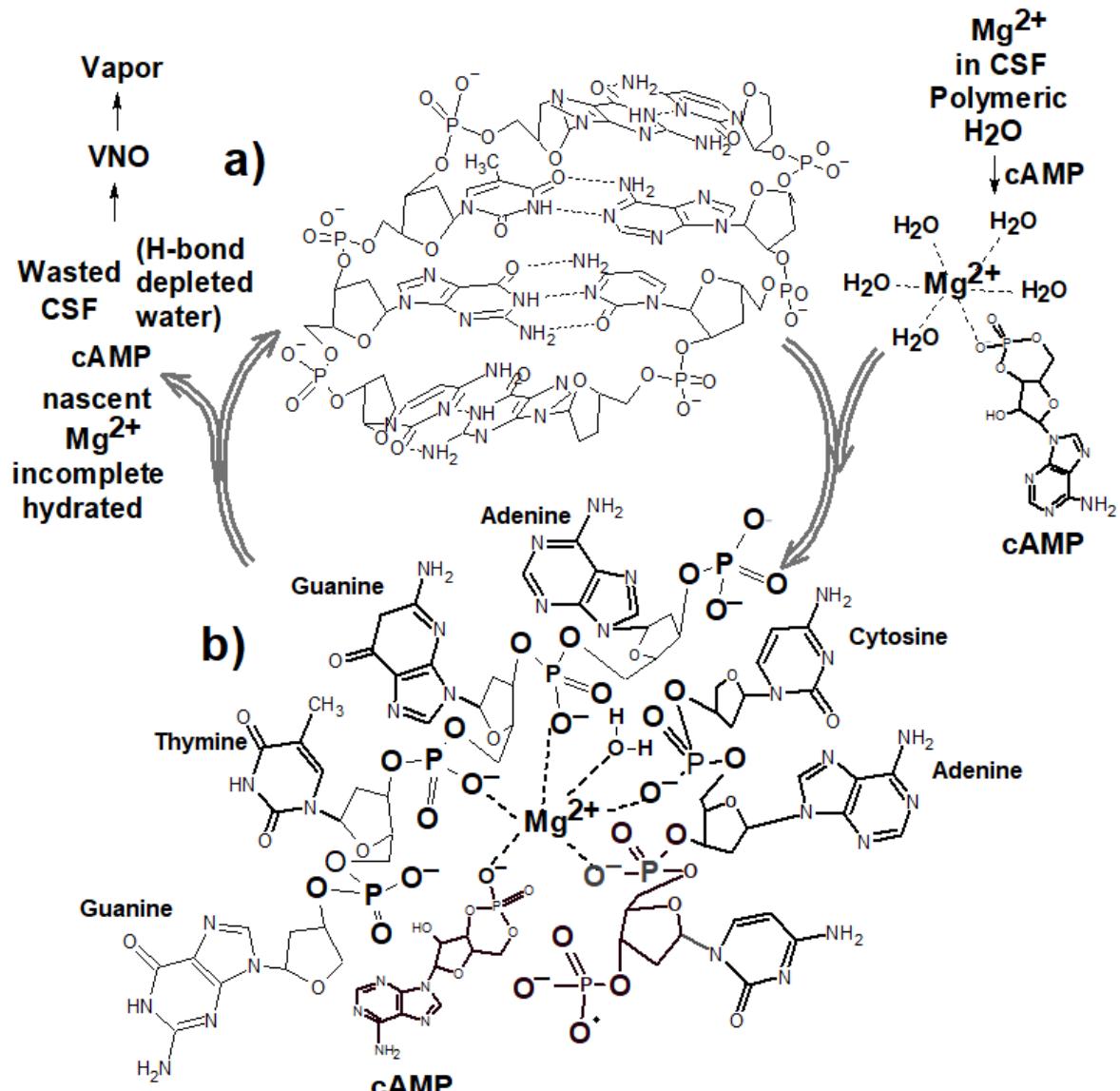


Figure 3. Physiological mechanism for chemical cAMP fitting into the double strands unzipping structure of cffDNA and localized for unzipping of a DNA. a) Base sequence of the two chains attracted to match in a double stranded binary rotational symmetry of DNA in the brain itself. b) cAMP unzipping mechanism opens the double-stranded DNA structure positioning to the outside purines and pyrimidines bases. Mg-cAMP binds to the pentahydrated Mg²⁺ ion to coordinate to both chains through the negatively oxygen of phosphate groups, on both of the backbone, connecting the repeated pattern of sugars and on that of cAMP [6, 14]. The DNA expression occurs by the transcription function. The triplet pairing rule for synthetizing mRNA and recognize their tRNAs partners are based upon each of the three bases pairing with its appropriate partner.

The non-physiological treatment technic of heating DNA at 65°C allows the strands separation and transcription that has been used experimentally.

The cAMP-Mg-DNA acts as a physiological process because can achieve a local opening of the double-stranded by the insertion of 3'-5' cyclicAMP through the cyclic configuration of its phosphoryl group negative charged oxygen, to face the hexahydrated Mg²⁺ and allowing the DNA chains to rotate for the purine and pyrimidine groups to face outwards.

The figure 3 shows that the phosphoryl groups of the opening in the DNA are now facing with their charged oxygen (O⁻) to the inside to bind coordinately to Mg²⁺ [31].

The catabolite activator protein (CAP) functions by binding in the presence of the allosteric promoters and enhances the ability of RNA polymerase holoenzyme (RNAP) to bind and initiate transcription [32].

The activation of Mg²⁺ stimulated adenylate cyclase results in cAMP production, which in the newborn cerebrospinal fluid (CSF) unzipping of the cell-free DNA (cfDNA) and cell-free fetal DNA (cffDNA), for mRNA production in the glial cells of cortical, hippocampal, and spinal cord. The responses to oxytocin, released from posterior pituitary results in the development of bonding memory.

The cffDNA is from the fetal vestige cells originated from a never configured olfactory bulb.

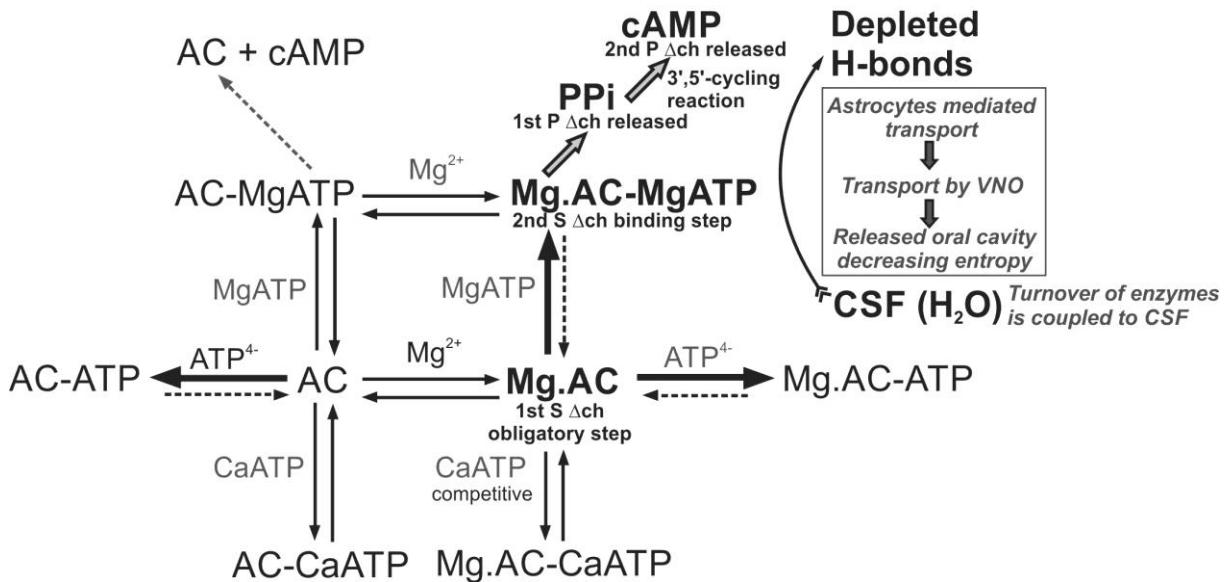
The cfDNA circulating in the maternal blood originates from cells shed from placental trophoblasts microparticles [33], disappears after two hours from delivery. Prenatal diagnosis targets on the cffDNA analysis show correspondence with the gene responsible for the sex-determining region Y protein (SRY) on the Y chromosome and the DYS14 sequence [34] [35].

Ionic equilibrium controlling Ca²⁺ effects for a simultaneous CaATP inhibition of adenylate cyclase and activation of the AMPA receptor

Drummond et al., 1971, Johnson and Sutherland, 1973, show that the responsiveness of AC to stimulation by adrenaline of fat-cell to noradrenaline(NA)-brain-AC-activated or basal hypothalamic-AC [13] is controlled by obligatory ions Mg²⁺ requirement exceeding participation in substrate formation. cAMP and Ca²⁺ signaling determine amplitude, phase and period of circadian rhythms [36]. Kinetic characterization of the hypothalamic adenylate cyclase response to Ca²⁺ releases to activate the glutamate neurotransmission. Serotonin (5-hydroxytryptamine, 5-HT) produced in Raphe nuclei located in the brainstem, could induced Ca²⁺ increase and reduced the cAMP increase, indicating cross-talk between the 5-HT-sensitive Ca²⁺ and cAMP pathways [37].

The responsiveness of AC to stimulation by hormones or neurotransmitters is controlled by an obligatory Mg²⁺ requirement that exceeds the one participating in substrate formation [38].

The finding that CF₁-ATPase requires Mg²⁺ for binding to the membrane and reconstitution of allotopic properties [39] suggests a similar obligatory participation of Mg²⁺ in the formation of a hormone receptor-Mg²⁺-AC complex [40]. Thus, the RARE BiBi mechanism predicts that hormones will increase *h*(MgCl₂) from basal =2.6 to 4, because ATP⁺ and AC are competitive ligands of Mg²⁺ to form an AC-Mg²⁺ and Mg²⁺-AC-MgATP complex. An NA additional AC saturation parameter adds to a higher *n* value. The experiment described here has been restricted to characterize the basal kinetics of the enzyme only, without hormone interaction with the hormone receptor an Mg²⁺-dependent integration with the membrane, to avoid a higher equilibria complexity.



Scheme 1. RARE BiBi (2 substrates and 2 products) ordered binding (macro mechanism) of adenylate cyclase including ATP⁴⁻ and CaATP as dead-end inhibitions. Applying the initial rate studies for Mg²⁺ could be assumed to be equally valid for Mn²⁺. S₁=Mg²⁺, S₂=MgATP; P₁=PPi (pyrophosphate); P₂=cAMP (3',5'-cyclic adenosine monophosphate); E=AC; Δch=conformational change. These are tests the close systems, but physiological acts the dissipative release out of the open system. If AC is inhibited the turnover disappears and the dissipative coupling of H-bonds also.

The equilibrium constants can be measured in a test, but the coupling of a dissipative H-bond reaction, favors non-equilibrium with continuous consumption of substrate and products, because the system becomes open. Such systems expel heat and entropy. However, Ca²⁺ inhibition of the AC enzyme prevents turnover and consumption of the H-bonds. Thus, coupling to other proteins turnover will persist.

Coupling by CSF loss of H-bonds by turnover of AC from hydrophilic to hydrophobic, transported by astrocytes and mediated by VNO is dissipated from the open system and allows entropy dissipation.

The RARE BiBi mechanism shows a second-order dependence on substrate concentration: Mg²⁺ has to bind first to activate the binding site for MgATP.

Thereafter a 3',5'-cyclic is formed, to produce cAMP and PPi are released. The sequential steps forming by conformational changes of obligatory binding sites for S₁, allowing S₂ binding, and specific conformational changes for 1st product, the release of PPi before cycling, and after release of 2nd product, cAMP.

The scheme 1 represents the molecular kinetics synchronization that prevents microscopic reversibility, because could not be conceptually assimilated as a single door, which could only allow transit in both senses. The scheme illustrates as a similitude to the entrance and exit points of a submarine tower two hatches: 1st, open and 2nd closed, enter and 1st closed and 2nd open. Thus, microscopic irreversibility results from the enzymes turnover between their hydrophilic and hydrophobic forms. Change of protein conformation turnover is supported by the activation energy of broken H-bonds, from polymeric water in cerebrospinal fluid (CSF), conversion into waste water, a depleted of coordinative bonds between molecules. Astrocytes could maintain the waste state of water in a liquid state until their release as vapor to the outside of the system, equivalent to entropy dissipation.

AC shows a RARE BiBi mechanism with death-end inhibitors of the active site: ATP⁴⁻ and Ca-ATP. Alternatively, a decrease in chelating metabolites (ATP⁴⁻) decreases CaATP, strongly activating AC-mediated and cAMP-dependent activation of pathways for memory affirmation. Turnover, with release of Mg²⁺ from the E as a nascent ion Mg²⁺ acquires a stronger intrinsic charge.

The effects of divalent metals and/or ATP⁴⁻ (in excess of their participation in complex formation) were determined from the corresponding apparent affinity values and the following kinetic constants were obtained: Mg²⁺ (Mg_T) can easily displace Ca²⁺ from ATP. The equilibrium favors that Mg²⁺ can easily displace Ca²⁺ from ATP, and reduce the concentration of CaATP. Since K_a

$(MgATP) = -20 \times 10^3 M^{-1}$, $K_i(ATP^4-) = 0.27 mM$, $K_a(CaATP) = -9 \times 10^3 M^{-1}$, $K_i(CaATP) = 0.015 mM$. ATP^4- and $CaATP$ were shown to compete for the active site of the enzyme.

Coupling of H-bonds consumption for proteins/enzymes turnover

The cerebrospinal fluid (CSF) expended H-bond water is in liquid state at $36.6^\circ C$ because the H-bond between molecules has been broken, allowing a transition state in which the internal (intrinsic) structure of the water molecule itself, has absorbed vibrational and rotational kinetic energy. This allows an aggregate state, until the space allows the translational energy that characterizes the vapor state. In physics the phenomenon is described as a transition state of second order that became independent of the microscopic structure. In a laboratory is well known that the distilled and condensed water is highly active (energy excess on the individual molecules) and has to be stationed for 24hs, before the fitting between water molecules allows their full H-bonding state.

Approaching a mirror to the mouth, a condensation test for vital signs, allows detection of a 5% vapor present in breath, to become evident. The thermodynamics turnover for an out of the system release of waste water, maintains a dissipative state characteristic of open systems. Thus, prevents a reversal of the metabolic flow and therefore conserve the energy capable to support the hydration shell turnover of ions and proteins, which maintains the cell membrane action potential.

The blood-brain capillaries release metabolites to the CSF interphase with glial cells. The toxic products into the CSF interphase with the venous system.

Astrocytes through the rapid circulation function as a radiator could prevent the brain could absorbing the entropy of the 30% the total calories ingested by the individuals.

Thermodynamic of cerebrospinal fluid (CSF) daily turnover

Thermodynamically a donor solvation media, like CSF could be calculated on the bases of a turnover value of 500ml CSF, which could be expressed as 27.77 H_2O mol, considering an average value of 2.3 mol H-bond per mol H_2O and -2.6kcal per mol H-bond.

$$Energy = 27.77 \text{ mol } H_2O \frac{2.3 \text{ mol H-bond}}{1 \text{ mol } H_2O} \frac{-2.6 \text{ kcal}}{\text{mol H-bond}} = -166 \text{ kcal}$$

Outside the body exhausted H-bond water regenerates by cooling into cluster water because is a favorable thermodynamic process.

The epithelial membranes with an outside and inside confers the properties of open systems, because the depleted H-bonds from water in CSF does not have the tendency to aggregate, but by entering in the spongy tissue of the palate it rapidly became separated in individual molecules and evaporate.

Thus, exhaled air in adults of about 6 liters per minute has a 5% vapor contribution from the VNO conductance process of depleted H-bonds from water in CSF. These show the conduction to entropy dissipation.

cAMP Mg-dependent zipping-out of DNA results in an cAMP- Mg^{2+} -DNA complex open up for regulation of cffDNA expression in the newborn, memory dependent plasticity, and eventually, a memory circuit in the growing up could be formed to operate for long-term memory. Erythrocytes are a carrier of cGMP to complete the cAMP signaling [41].

The hydration shell of *nascent* Mg^{2+} allows capture molecules of water from the hydration sphere of Na^+ and this one in term replaces this loss from capture of H_2O from the hydration sphere of K^+ . The sequence allows the sieve effects, required to activate the electrogenic pump and the potential of neuronal membrane.

Not much progress has preceded this report on the complementary structure of the cAMP-Mg-DNA complex for unzipping of the double helix, ignoring that there is a need to relate the well-known effect of cAMP of memory affirmation pathways for structuring neuronal circuits.

Mg^{2+} has been shown required for specific binding of the double-stranded DNA containing the consensus CRE sequence (figure 3).

The product of AC activation is cAMP which correlates with the central role of the enzyme subtract. Thus, the reactions of cAMP with Mg²⁺ and water result in unzipping DNA through the formation of Mg²⁺-cAMP-DNA complex. The cAMP-response-element-binding (CREB) was proposed in 1987 as a cAMP-responsive transcription factor regulating the somatostatin gene [42].

NA (noradrenaline) released by the long axons the synaptic junctions of the corpus coeruleus neurons, also contributes to the additional regulation of AC, connecting brain regions to the same event, reaching a more comprehensive understanding.

Discussion

Central role of cAMP for the regulation and integration hypothalamic response and memory through DNA unzipping

Oxytocin produced by the hypothalamus, stored and secreted by the posterior pituitary gland acts as a neurohypophysial/neuromodulator of intimacy, sex and reproduction. It is released in large amounts after distension of the cervix and uterus during labor, facilitating birth, maternal bonding, and, after stimulation of the nipples, lactation [43].

The locus coeruleus neurons by the effect of NA acting on the brain led to the recognition of the molecular equivalent for the psychosomatic axis and its impact on many social problems.

A mechanism separating emotional-long-term (LTM) from short-term memories (STM) also called working memory may not require totally independent pathways. Both could operate from CREB mediate process of accumulation of transduced information into peptides. The latter, could be selectively encoded. Natural selection may not ascribe long-term neural patterns, based on parenting experiences, to later allow a more comprehensive brain reorganization as an adult and avoid the animal compulsions of a reflex conditioning described by Pavlov, which in humans if present as compulsive will be restricted by evolution of social impact.

Genes are involved in the synthesis of oxytocin peptides and other hormones, which promote reflection in humans, allowing cultural adaptation, which operates to control the responses of humans to instinctive secretion of hormones, which require social repression such as behavior of femicide.

Working memory could be expressed because emotional conditions lead to stimulate constitutive gene expression of oxytocin recognition receptors. Oxytocin could be measured in saliva samples. At the spines and/or dendrites, the binding of Mg²⁺ stimulates the oxytocin-occupied receptor for a rapid release of the pheromone response.

Social olfactory deficits in mice without the oxytocin gene [44] [45] are rescued with injection of oxytocin into rat olfactory bulbs. Oxytocin-mRNA has been shown to be highly expressed in human paraventricular nucleus of the hypothalamus, the lateral hypothalamic area, and the supraoptic nucleus [46].

Increased mRNA in the human olfactory epithelium region may be vestigial, as olfaction is not as important for human conspecific identification, compared to most other mammals due to species specialization.

Oxytocin's feedback secretion is controlled by Ca²⁺ release expected to be a potential therapeutic resource for the social core symptoms of autism spectrum disorder (ASD), since this neuropeptide can modulate human social behavior and cognition [47]. Oxytocin interacts with the neural pathways responding to motivationally relevant stimuli and dopamine system [48] [49] [50] [51] [52], craving for oxytocin induces aggressive response to abandonment like gender violence [6].

Dopamine's feedback through the connection of the portal system with the tongue plexus sponge tissue is a self-reward for many risks deports, conditioning the unconscious and conscious level, which is essential to voluntary motion and cognition. This improves the results of treatments in newborns with suspected lack of oxygen during childbirth, also regulates motivation and the reward system, related to love through desire.

H-bonds thermodynamics contribution to dissipative states of the open system

A model of mutual inclusion operates during transition of the Oxy- to Deoxy- forms of Hb. Thus, conformational dynamics for its hydro- to dehydro- forms, could release O₂ and Mg²⁺ for a flow of matter and energy, during the function the arterial irrigation of the brain's blood-astrocyte-neuronal system.

The kinetics of brain adenylate cyclase (AC) have a high basal activity that analyzed by a RARE BiBi mechanism (scheme 1) shows an obligatory excess of Mg²⁺ (S₁), over Mg-ATP (S₂) prior to the binding of S₂, indicating that Mg²⁺ is required to configure a site for MgATP binding.

Turnover of the enzyme release Mg²⁺ from the E as a nascent ion (n-Mg²⁺), which shows a stronger intrinsic charge. Microscopic reversibility is restricted by coupling the cerebrospinal fluid (CSF), containing polymerized water to the loss of H-bonds that eventually are released as vapor, as an entropy decrease mechanism.

A GTP cycle is coupled to noradrenaline (NA) active site. Adrenaline is not present in brain; it has separated functions like an increased heart rate and preparing the body for stress [1]. NA is released by the long axons of the 8.6x10⁴ neurons of the locus coeruleus into the synaptic junctions activating AC for sensorial-integrated perception areas for an all-inclusive memory function. Because of the blood-brain barrier (BBB), noradrenaline play separated roles for brain AC that adrenaline for the body AC.

The lack of adrenaline access by the BBB prevents a negative feedback for metabolic homeostatic control of the hypothalamic–pituitary–adrenal (HTPA) axis. Thus, allowing persistence of stress-stimulus, acting on the hypothalamus. This increases the release of orexins A and B. Orexin-A is capable to stimulate the adrenocortical cells to secrete cortisol (figure 2). This one leads to a constant increase in cortisol, and thus becoming the main stress-hormone.

The activation of the Na⁺/K⁺-ATPase pump, by decreasing ATP⁴⁻, release the ion Mg²⁺ with an activatory effect on AC.

Mg-cAMP and Mg-cGMP (carried by erythrocytes) controls the physiological expression of brain DNA. These are expected to contribute to the understanding of plasticity, and its relationship to the response to learning, by creating neuronal circuits, which characterize memory. Stress mediated by increasing adrenaline secretion, usually shows a pattern of hyperfunction follow by hypofunction, which when the stressed organ is brain, it could be associated with a symptomatology of persistent anxiety followed by depression [1].

Under prolonged stress, the synthesis of new AC, may not match the rate of NA-dependent AC inactivatory decrease. The viability of functional synapses may be controlled by the speed of synthesis of the proteins, which make-up the hormonal receptor sites and/or AC-itself, versus the rate of inactivation or destruction of AC.

A restriction on the randomness of evolution could result, from sensorial signaling for self-dependent response evolution to pheromone or saliva hormonal communication by the G-protein 7TM receptors, activating hypothalamic AC. The cAMP produced could form cAMP-Mg-cffDNA complex transcribed in the messenger RNA, modulating brain plasticity by acting as a feedback on brain evolution, capable to overcome either the randomness, or inherited patterns, but adapting the developing neuronal circuit network for a greater cognitive capacity and social-family.

Conclusions

Cell-free fetal DNA (cffDNA) appears as an evolutionary disaggregation of the olfactory bulb. Hypothalamic cAMP unzipping DNA for transcription as RNA messenger acts as a carrier of emotional needs of the unconscious olfactory sensorial axis, during lactation. Human brain function has a constitutive and functional separation of integrated parameters: the one for synapsis and neuronal circuits, from that of a multiple wrapped-around astrocytes and their exchanges. The brain operates as a thermodynamic open system for the summation of the energy generated by metabolites and H-bond consumption. Thus, supports the potential of the dissipative states above their equilibrium ratios of enzyme substrate and product concentrations, and the electrogenic action potentials. The self-reward emotional feedback could differentiate animically most persons, which at

the psychoanalytic level will manifest tendencies of dissimilar social fitness. Contributions when could be modified by literacy as showed differentiable interconnectivity or brain areas. The latter, will restrict the randomness of brain evolution, allowing cerebral self-evolving improvement of affective links and social influences, affecting brain plasticity.

The AC up-regulation by Mg^{2+} is turning off by Ca^{+2} . The stressors trigger the Mg^{2+} response and subsequently by an excess of free Mg^{2+} over subtract activates AC unchaining the fight or flight response.

The regenerative capacity of cAMP could be used for treatment in Alzheimer's disease could be selected in order to reach specific brain areas. It could be assumed that dysfunctions of the vomeronasal pathway during nurturing could lead to autism.

Excitotoxicity due to excessive glutamate release and impaired uptake occurs as part of the ischemic cascade and is associated with stroke [53], autism [54], some forms of intellectual disability, and diseases such as amyotrophic lateral sclerosis, lathyrism, and Alzheimer's disease [55]. In contrast, decreased glutamate release is observed under conditions of classical phenylketonuria [56] leading to developmental disruption of glutamate receptor expression [57].

It is not known how adrenaline could possible reach CSF but if does may be one of the causals for schizophrenia, because the disease is known to show disruption of feeding-starvation process [58].

Link to Rutgers Library: https://www.researchgate.net/profile/Alfred_Bennun

Open access to full-texts publications and 3 books.

References

- [1] Brydon-Golz, S. and Bennun, A. (1975). Postsynthetic stabilized modification of adenylate cyclase by metabolites. *Biochemical Society Transactions*, 3, 721-724.
- [2] Gravina S, Sedivy JM, Vijg J (June 2016). The dark side of circulating nucleic acids. *Aging Cell*. 15 (3): 398–9.
- [3] Trotier, D. (2011) Vomeronasal organ and human pheromones. *European Annals of Otorhinolaryngology, Head and Neck Diseases*. Volume 128, Issue 4, Pages 184-190.
- [4] Gray's Anatomy: The Anatomical Basis of Clinical Practice (41 ed.). Elsevier Health Sciences. 2015. p. 358.
- [5] Yang HP, Wang L, Han L, Wang SC (2013). Nonsocial functions of hypothalamic oxytocin. *ISRN Neuroscience*. 2013: 179272.
- [6] Bennun A. The Metabolic-Psychosomatic Axis, Stress and Oxytocin Regulation Nova Publishers (2016) Serie: Biochemistry and molecular biology in the post genomic era.
- [7] Dulac, C. and Axel, R. (1995). A novel family of genes encoding putative pheromone receptors in mammals. *Cell*. 83 (2): 195–206.
- [8] Matsunami, H. and Buck, L.B. (1997) A multigene family encoding a diverse array of putative pheromone receptors in mammals. *Cell*. 90(4):775-84.
- [9] Ryba, N.J. and Tirindelli, R. (1997). A new multigene family of putative pheromone receptors. *Neuron*. 19 (2): 371–9.
- [10] Rivière, S.; Challet, L.; Fluegge, D.; Spehr, M. and Rodriguez, I. (2009). Formyl peptide receptor-like proteins are a novel family of vomeronasal chemosensors. *Nature*. 459 (7246): 574–7.
- [11] Liberles, S.D.; Horowitz, L.F.; Kuang, D.; Contos, J.J.; Wilson, K.L.; Siltberg-Liberles, J.; Liberles, D.A. and Buck, L.B. (2009). Formyl peptide receptors are candidate chemosensory receptors in the vomeronasal organ. *Proceedings of the National Academy of Sciences of the United States of America*. 106 (24): 9842–7.
- [12] Rivière, S.; Challet, L.; Fluegge, D.; Spehr, M. and Rodriguez, I. (2009). Formyl peptide receptor-like proteins are a novel family of vomeronasal chemosensors. *Nature*. 459 (7246): 574–7.
- [13] Ohanian, H.; Borhanian, K.; De Farias, S. and Bennun, A. (1981) A model for the regulation of brain adenylate cyclase by ionic equilibria. *Journal of Bioenergetics and Biomembranes*, Vol. 13, 5/6, 317-55.
- [14] Vicario, P. P.; Saperstein, R. and Bennun, A. (1988). Role of divalent metals in the kinetic mechanism of insulin receptor tyrosine kinase. *Arch. Biochem. Biophys.*, 261(2), 336-45.
- [15] Bennun A. (2012). Molecular Mechanisms Integrating Adenylyl Cyclase Responsiveness to Metabolic Control on Long-Term Emotional Memory and Associated Disorders. Nova Science Publishers, Inc. Long-Term Memory: Mechanisms, Types and Disorders (1-44). New York, USA.
- [16] Barchi, R. L. (1998). Ion channel mutations affecting muscle and brain. *Curr Opin Neurol.*, 11(5), 461-8.
- [17] Sun, W.; Barchi, R. L. and Cohen, S. A. (1995). Probing sodium channel cytoplasmic domain structure. Evidence for the interaction of the rSkM1 amino and carboxyl termini. *J. Biol. Chem.*, 270 (38), 22271-6.
- [18] Moran, D.T., Jafek, B.W. and Rowley, J.C. (1991) The vomeronasal (Jacobson's) organ in man: ultrastructure and frequency of occurrence. *J. Steroid Biochem. Mol. Biol.*., 39, 545–552.
- [19] Stensaas, L.J.; Lavker, R.M.; Monti-Bloch, L.; Grosser, B.I. and Berliner, D.L. (1991) Ultrastructure of the human vomeronasal organ. *J. Steroid Biochem. Mol. Biol.*., 39(4B), 553–560.
- [20] Han, X., et al. (2013) Forebrain engraftment by human glial progenitor cells enhances synaptic plasticity and learning in adult mice. *Cell Stem Cell* 12, 342–353.
- [21] Herculano-Houzel, S. (2014) The glia/neuron ratio: how it varies uniformly across brain structures and species and what that means for brain physiology and evolution. *Glia* 62, 1377–1391.
- [22] Kimelberg, H.K. and Nedergaard, M. (2010) Functions of astrocytes and their potential as therapeutic targets. *Neurotherapeutics*. 7, 338–353.
- [23] Oberheim, N.A., et al. (2009) Uniquely hominid features of adult human astrocytes. *J Neurosci* 29, 3276–3287.
- [24] Iadecola, C. and Nedergaard, M. (2007) Glial regulation of the cerebral microvasculature. *Nat.Neurosci.* 10, 1369–1376.
- [25] Simard, M. and Nedergaard, M. (2004) The neurobiology of glia in the context of water and ion homeostasis. *Neuroscience* 129, 877–896.
- [26] Silva, A. J.; Kogan, J. H.; Frankland, P. W. and Kida, S. (1998). CREB and memory. *Annu. Rev. Neurosci.*, 21, 127-48.
- [27] Davis, H. P. and Squire, L. R. (1984) Protein synthesis and memory: a review. *Psychol. Bull.*, 96 (3), 518-59.
- [28] Mayr, B. and Montminy, M. (2001) Transcriptional regulation by the phosphorylation dependent factor CREB. *Nat. Rev. Mol. Cell Biol.*, 2 (8), 599-609.

- [29] Kandel, E. R. (2012). The molecular biology of memory: cAMP, PKA, CRE, CREB-1, CREB-2, and CPEB. *Mol. Brain.*, 5, 14.
- [30] Kandel, E. R. (2001). The molecular biology of memory storage: a dialogue between genes and synapses. *Science*, 294 (5544), 1030-8.
- [31] Moll, J. R.; Acharya, A.; Gal, J.; Mir A. A. and Vinson C. (2012). Magnesium is required for specific DNA binding of the CREB B-ZIP domain. *Nucleic Acids Res.*, 30(5), 1240-6.
- [32] Lawson, C. L.; Swigon, D.; Murakami, K. S.; Darst, S. A.; Berman, H.M. and Ebright, R. H. (2004). Catabolite activator protein: DNA binding and transcription activation. *Curr Opin Struct Biol.*, 14(1), 10-20.
- [33] Smets, E.M.; Visser, A.; Go, A.T.; van Vugt, J.M. and Oudejans, C.B. (2006) Novel biomarkers in preeclampsia. *Clin Chim Acta*. 364(1-2):22-32.
- [34] Bustamante-Aragones, A.; Gonzalez-Gonzalez, C.; de Alba, M.R.; Ainse, E. and Ramos, C. (2010) Noninvasive prenatal diagnosis using cfDNA in maternal blood: state of the art. *Expert Review of Molecular Diagnostics*. Informa UK Limited., 10 (2): 197–205.
- [35] Zimmermann, B.; El-Sheikhah, A.; Nicolaides, K.; Holzgreve, W. and Hahn, S. (2005) Optimized real-time quantitative PCR measurement of male fetal DNA in maternal plasma. *Clin Chem*. 51(9):1598-604.
- [36] O'Neill, J.S. and Reddy, A.B. (2012) The essential role of cAMP/Ca²⁺ signalling in mammalian circadian timekeeping. *Biochem Soc Trans.* 40(1): 44–50.
- [37] Amireault, P. and Dubé, F. (2005) Intracellular cAMP and calcium signaling by serotonin in mouse cumulus-oocyte complexes. *Mol Pharmacol*. 68(6):1678-87.
- [38] Harris, R.; Cruz, R. and Bennun, A. (1979). The effect of hormones on metal and metal-ATP interactions with fat cell adenylate cyclase. *BioSystems*, 11, 29-46.
- [39] Bennun, A. and Racker, E. (1969) Partial resolution of the enzymes catalyzing photophosphorylation IV. Interaction of coupling factor I from chloroplast with components of the chloroplast membrane. *J. Biol. Chem.*, 244, 1325-1331.
- [40] Harris, R. and Bennun, A. (1976). Hormonal control of fat cells adenylate cyclase. *Molecular & Cellular Biochemistry*, 13 (3), 141-146.
- [41] De Bari, V.A. and Bennun, A. (1982) Cyclic GMP in the human erythrocyte. Intracellular levels and transport in normal subjects and chronic hemodialysis patients, *Clinical Biochemistry* 15(4), 219-221.
- [42] Montminy, M.R. and Bilezikian, L.M. (1987) Binding of a nuclear protein to the cyclic-AMP response element of the somatostatin gene. *Nature*, 328 (6126): 175–178
- [43] Marieb, E. N. and Hoehn, K. N. (2012) Human Anatomy & Physiology 9th edition, chapter:16, page:599. Series: Books a la Carte. Publisher: Pearson.
- [44] Levy, F.; Kendrick, K.; Goode, J.; Guevara-Guzman, R. and Keverne, E. (1995) Oxytocin and vasopressin release in the olfactory bulb of parturient ewes: changes with maternal experience and effects on acetylcholine, γ-aminobutyric acid, glutamate and noradrenaline release. *Brain Res.* 669, 197–206.
- [45] Ferguson, J. N. et al. (2000) Social amnesia in mice lacking the oxytocin gene. *Nat. Genet.* 25, 284–288.
- [46] Dluzen, D. E.; Muraoka, S.; Engelmann, M. and Landgraf, R. (1998) The effects of infusion of arginine vasopressin, oxytocin, or their antagonists into the olfactory bulb upon social recognition responses in male rats. *Peptides* 19, 999–1005.
- [47] Yamasue, H. and Domes, G. (2018) Oxytocin and Autism Spectrum Disorders. *Curr Top Behav Neurosci*. 35:449-465.
- [48] Love, T.M. (2014) Oxytocin, motivation and the role of dopamine. *Pharmacol Biochem Behav.*, 119:49-60.
- [49] Bennun A. The Regenerative Processes Involving the cAMP Unzipping of DNA. The Synthesis of Proteins Integrating Plasticity and Longevity. *Biochemistry Research Trends*. Book Published by Nova Biomedical, Copyright 2017 by Nova Science Publishers, Inc.
- [50] Bennun A. The assay of the hydration shell dynamics on the turnover of the active site of CF1-ATPase. Book Title: Advances in Chemistry Research. Volume 33, chapter 8. Editor James C Taylor. Nova Publishers (2016).
- [51] Bennun A. The integrated model of cAMP-dependent DNA expression reveals an inverse relationship between cancer and neurodegeneration. Book Title: Horizons in Cancer Research. Volume 63, Chapter 9 (Book ID: _10383_). Nova Publishers (2016)
- [52] Bennun A. The Noradrenaline-Adrenaline-Axis of the Fight-or-Flight Exhibits Oxytocin and Serotonin Adaptive Responses. *International Journal of Medical and Biological Frontiers*. Volume 21, Issue 4, pages: 387-408 (2015).
- [53] Sapolsky, R. (2005) Biology and Human Behavior: The Neurological Origins of Individuality, 2nd edition. The Teaching Company. see pages 19 and 20 of Guide Book.
- [54] Shinohe, A.; Hashimoto, K.; Nakamura, K.; Tsujii, M.; Iwata, Y.; Tsuchiya, K.J.; Sekine, Y.; Suda, S.; Suzuki, K.; Sugihara, G.; Matsuzaki, H.; Minabe, Y.; Sugiyama, T.; Kawai, M.; Iyo, M.; Takei, N. and Mori, N.

- (2006) Increased serum levels of glutamate in adult patients with autism. *Progress in Neuro-Psychopharmacology & Biological Psychiatry*. 30 (8): 1472–7.
- [55] Hynd, M.R.; Scott, H.L. and Dodd, P.R. (2004) Glutamate-mediated excitotoxicity and neurodegeneration in Alzheimer's disease. *Neurochemistry International*, 45 (5): 583–95.
- [56] Glushakov, A.V.; Dennis, D.M.; Sumners, C.; Seubert, C.N. and Martynyuk, A.E. (2003) L-phenylalanine selectively depresses currents at glutamatergic excitatory synapses. *Journal of Neuroscience Research.*, 72 (1): 116–24.
- [57] Glushakov, A.V.; Glushakova, O.; Varshney, M.; Bajpai, L.K.; Sumners, C.; Laipis, P.J.; Embury, J.E.; Baker, S.P.; Otero, D.H.; Dennis, D.M.; Seubert, C.N. and Martynyuk, A.E. (2005) Long-term changes in glutamatergic synaptic transmission in phenylketonuria. *Brain*, 128 (Pt 2): 300–7.
- [58] Mackliff, J. R. Schizophrenia and Parkison surgery: A new and efficient regulation of dopaminergic synapses after B.E.A.M. (Bilateral Electrocoagulation of Adrenal Medulla). Editorial: Amazon (2017) ISBN: 1539387887.