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Review Article

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Autophagic processes of normal and malignant metabolic pathways

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Abstract

Autophagy is a cellular process that maintains the homeostasis of the normal cell, but autophagic dysfunction is associated with human diseases, such as cancer. In normal cells, the initial signal to form auto-phagosomes is by the class III phosphatidyl-inositol (PI) 3 kinase complex consisting of sequence genes, Beclin1/ Atg6 and class III PI3K (Vps34). This process is negatively regulated by binding of BcI-2 family members such as BcI-xL to Beclin1 preventing Beclin1 binding to the PI3K-III complex and thereby reducing autophagy. In cancer, the autophagy can be neutral, tumor-suppressive, or tumor-promoting in different contexts. Genomic analysis of human cancers indicates that the loss or mutation of core autophagy Atg genes, whereas oncogenic events that activate autophagy and lysosomal biogenesis have been identified in malignant diseases. Optimal combination of inductors or inhibitors of autophagy with chemo or radiotherapy in a variety of tumor type, in different phases, can be successful approaches for improve the effect of anticancer therapies.

Keywords: Autophagy, Apoptosis, BcI-2-protein, Phosphatidyl-inositol (PI) 3 kinase complex, Mammalian target of rapamycin (mTOR), Tensin-homologous phosphatase (PTEN).

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1. Introduction

Each cell is "programmed" to "commit suicide," and this happens every day, without feeling any affection and without your life being jeopardized. The "suicide" of cells is part of the sustainability of life on earth, so they die for the good of our body, they are the "heroes" that are found in each of us. In fact, in the absence of this mechanism, life would suffer.

Aging is not properly considered a disease; however, it is associated with different pathological conditions. In the last period of human life, cells undergo several changes, including DNA mutations, damages at several other molecules, and accumulation of protein aggregates. Several studies have demonstrated that autophagy activation protects from aging. In fact, not only autophagy levels decrease with age but also overexpression of autophagy-related genes (Atg) proteins contributes to improve life span in a human model of aging in vitro and in mouse models in vivo (Madeo *et al.*, 2015).

2. Roles of Autophagy in Human Pathology

As already mentioned, one of the most important functions of autophagy is the degradation of misfolded proteins, so in neurons, the failure of autophagy can contribute to neurodegeneration (Muller *et al.*, 2017). It

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can occur in Parkinson's disease, a neurodegenerative disorder characterized by α -synuclein accumulation in the brain. Recently, in a study, was demonstrated that autophagy inhibition by 3-methyladenine (3-MA) and by Atg5 knocking down in lymphocytes lead to a significant increase of α -synuclein levels (Colasanti *et al.*, 2014). The removal of mitochondria, source of Reactive Oxygen Species (ROS), performed by autophagy, certainly protects cells from DNA mutations and prevents cellular transformation. It has also been demonstrated that deletion of the autophagic gene Beclin-1 may cause development of various malignancies in mouse models (Galluzzi *et al.*, 2015).

Furthermore, it has been demonstrated that autophagy-deficient tumors are more sensitive to several chemotherapeutic agents (Levy *et al.*, 2014). In this case, autophagy promotes the survival of cancer cells and protects them from the action of drugs that induce apoptosis. Although research in this field is just at the beginning, an encouraging number of works suggest that defects in the autophagy mechanism may be involved in the pathogenesis of autoimmune diseases.

In Systemic Lupus Erythematosus (SLE), showed that factors present in the serum of SLE patients, probably antibodies, are able to induce autophagy in T lymphocytes from healthy donors, but not in T lymphocytes from patients with SLE. We speculated that chronic exposure to specific autoantibodies, as occurs in SLE, could lead to the selection of autophagy-resistant T lymphocytes (Alessandri *et al.*, 2012; and Barbati *et al.*, 2015).

3. Physiological Functions and Molecular Mechanism of Autophagy

Autophagy is a degradation pathway characterized by the isolation of targeted cytoplasmic material in a typical double-membrane vesicle, known as autophagic vacuole or auto phagosome (Ohsumi, 2014). The subsequent fusion of the auto phagosome with the lysosome ensures the correct destruction of organelles, misfolded proteins, and microorganisms, carried inside the vesicle. Despite its emerging role in human pathology, autophagy is a physiological process involved in basal organelles turnover and in the removal of proteins aggregates (Fougeray and Pallet, 2015), (Figure 1).

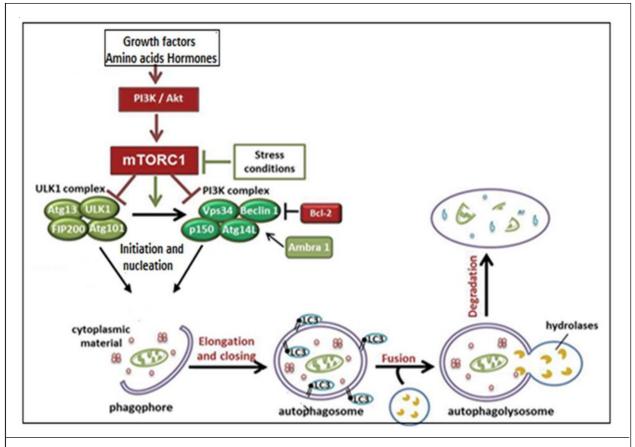


Figure 1: Schematic representation of signaling pathways involved in autophagy multistep regulation. In presence of growth factors and hormones, mTORC1 inhibits autophagy activation (Vomero M, Barbati C, Colasanti T, Perricone, C Novelli L, Ceccarelli F *et al.* Autophagy and Rheumatoid Arthritis: Current Knowledges and Future Perspectives. Front Immunol. 2018; 9: 1577. Published online 2018 Jul 18. doi: 10.3389/fimmu.2018.01577)

In response to the condition of cellular stress, such as growth factors and nutrients deprivation, intracellular components degraded by autophagy are recycled in order to generate ATP and sustain essential cell functions (Yu *et al.*, 2018). Autophagy is considered a pro-survival mechanism, allowing cells to respond to injury by degrading unnecessary and dysfunctional self-components; however, this ability may become a double-edged sword. Three types of autophagy can be distinguished: macro-autophagy, micro-autophagy, and chaperone-mediated autophagy. In this review, we will focus on macro-autophagy (hereafter referred to as autophagy), which is the most characterized type of autophagy.

Considering the crucial role of autophagy in the maintenance of cellular homeostasis, it is not surprising that several signaling-related molecules are involved in the perfect functioning of this process. Genetic screens in yeasts allowed the discovery of at least 37 autophagy-related genes (Atg) (Klionsky *et al.*, 2016). Many of these genes, encoding proteins involved in autophagy and its regulation, are evolutionarily conserved in humans.

The mammalian target of rapamycin (mTOR) complex 1 (mTORC1) regulates the activation of autophagy machinery, acting as a sensor of energy levels and integrating upstream signals deriving from other pathways, including the phosphoinositide 3-kinase (PI3K). In the presence of amino-acids and growth factors, mTORC1 represses autophagy by inhibition of Vps34 and ULK1 complexes. On the contrary, in low nutrients state, defined as starvation, the dissociation of mTORC1 from the induction complex triggers autophagy (Vomero et al., 2018; Yuan and Cantley, 2008; and Hosokawa et al., 2009), (Figure 2).

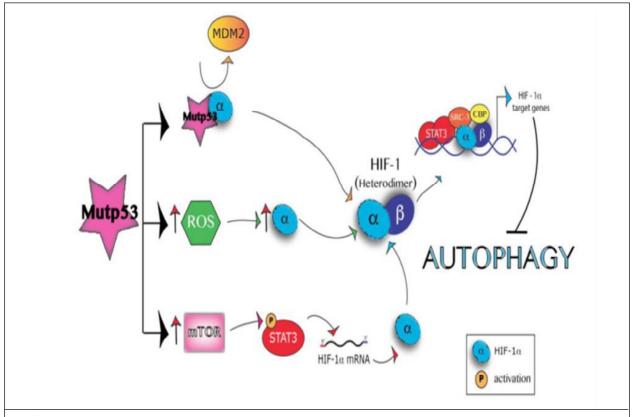


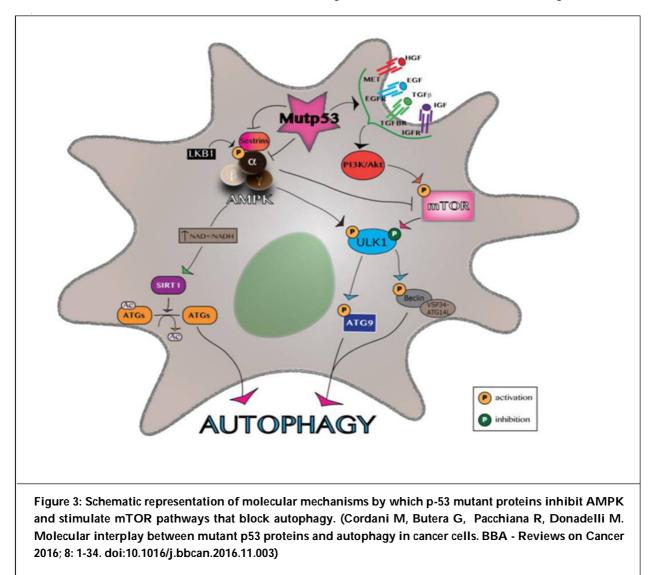
Figure 2: Schematic representation of the mechanisms at the basis of the stimulation of HIF-1 and HIF1- target genes by mutp53 resulting in autophagy inhibition. Regulation of HIF-1 α pathway at different levels. (a) Growth factors related pathways; (b) pVHL related pathways; (c) FIH-1 pathway; (d) Mdm2-p53 mediated ubiquitination and proteasomal degradation pathway. (Cordani M, Butera G, Pacchiana R, Donadelli M. Molecular interplay between mutant p53 proteins and autophagy in cancer cells. BBA - Reviews on Cancer 2016; 8: 1-34. doi:10.1016/j.bbcan.2016.11.003)

The autophagosome derives from a double-membrane pre-autophagosome structure called phagophore, which seems to originate from different sources, including plasma membrane (Ravikumar *et al.*, 2010), endoplasmic reticulum (YIa-Anttila *et al.*, 2009) and Golgi complex, in mammalian cells (Takahashi *et al.*, 2011). Phagophore nucleation requires the activity of class III phosphatidylinositol 3-kinase (PI3K-III) complex containing Beclin-1 (O'Farrell *et al.*, 2013).

The initial signal to form auto-phagosomes is by the class III phosphatidyl-inositol (PI) 3 kinase complex consisting of sequence genes, Beclin1/Atg6, p150hVSp35, and class III PI3K (Vps34). The autophagy promoting function of Beclin-1 is influenced by the antiapoptotic protein Bcl-2; in fact, when Beclin-1 is bound to Bcl-2, autophagy is inhibited; instead, the dissociation from Bcl-2 allows Beclin-1 to interact with PI3K-III complex, and to activate autophagy (Levine *et al.*, 2008). On the contrary, Beclin-1 regulated autophagy protein 1 (AMBRA1) is a positive regulator of Beclin-1-dependent autophagy; thanks to its capacity to create a link between cytoskeletal motor proteins and class III PI3K complex (Fimia *et al.*, 2007; and Di Bartolomeo *et al.*, 2013). Two ubiquitin-like conjugation systems, Atg12–Atg5–Atg16L and microtubule-associated protein 1 light-chain 3 (LC3)–phosphatidylethanolamine (PE), mediate the second step of autophagy, which concerns the expansion and closure of the auto phagosome (Tang *et al.*, 2015).

4. Protein p53-mediated Autophagy: A Prospective Strategy for Cancer Therapy

The regulation of autophagy is quite intricate. It involves a series of signaling cascades including p53, known as the best-characterized tumor suppressor protein. Recent reports have indicated that p53 plays dual roles in regulating autophagy depending on its subcellular localization. Consistent with this role, p53 activity is compromised in a high proportion of all cancer types, either through mutation of the TP53 gene (encoding p53) or changes in the status of p53 modulators. Phosphorylation of the N-terminal end of p53 by the specific protein kinase disrupts Mdm2-binding and activates p-53 protein. Stimulation of HIF-1 and HIF1- target genes by mutant p53 protein results in autophagy inhibition. Thus, autophagy defects impair survival of apoptosis-defective tumor cells upon nutrient and oxygen limitation, leading to cell death by necrosis, which in turn is associated with inflammatory cell recruitment, cytokine production and nuclear factor κ B (NF κ B) activation, which has been linked to accelerated tumor growth (Masoud and Wei Li, 2015), (Figure 3).



Integrity checkpoint, a molecular cascade that detects and responds to several forms of DNA damage caused by genotoxic stress. Oncogenes also stimulate p53 activation, mediated by the protein p14ARF, (Feng et al., 2015). In carcinogenesis are involved a series of signaling cascades including p53, known as the best-characterized tumor suppressor protein (Udristioiu and Nica-Badea, 2018). Recent reports have indicated that p53 plays dual roles in regulating autophagy depending on its subcellular localization. Nuclear p53 facilitates autophagy by trans activating its target genes, whereas cytoplasmic p53 mainly inhibit.

Many cancer cells contain inactivating mutations of p53, which explains why those cancer cells go on living (Tasdemir *et al.*, 2008). The tumor suppressor p53 is a critical checkpoint protein in mammalian cells which is activated under genotoxic stress conditions, including DNA damage, hypoxia and oncogene activation, and responds by initiating tumor suppression mechanisms, such as cell cycle arrest, senescence and apoptosis. Under these conditions, p53 has been shown to trans activate autophagy-inducing genes and stimulate autophagy by inhibiting mTOR in an AMP-activated protein kinase (AMPK) (Feng *et al.*, 2007). Thus, positive regulators of apoptosis also induce autophagy, which is not very surprising given that both pathways are activated under similar stress conditions (Tracy *et al.*, 2007).

On the other hand, when autophagy is blocked, apoptosis is accelerated or apoptosis-defective cells undergo metabolic catastrophe and die by necrosis. Thus, cell fate in response to metabolic stress is determined by the functional status and the interaction between the stress-mitigating pathways of apoptosis and autophagy (Boya *et al.*, 2005; and Jin *et al.*, 2007). Regarding cancer prevention, the function of autophagy as a protector of cellular homeostasis and genome integrity may be particularly important. Autophagy inhibition as a means to sensitize cancer cells to treatment has been validated in several studies: inhibition of autophagy by chloroquine, a lysosome-tropic agent that raises intra-lysosomal pH and interferes with auto phagosome degradation within lysosomes, was shown to enhance the anticancer activity of the alkylating agent cyclophosphamide in a myc gene. Gene-induced lymphoma model and to induce p53-dependent cell death and tumor suppression in different myc-induced and ATM-deficiency lymphoma models (Amaravadi *et al.*, 2007).

5. Role and Regulation of Autophagy in Cancer

Increasing evidence reveals that autophagy dysfunction is associated with human diseases, such as cancer. Paradoxically, although autophagy is well recognized as a cell survival process that promotes tumor development, it can also participate in a caspase-independent form of programmed cell death. Induction of autophagy cell death by some anticancer agents highlights the potential of this process as a cancer treatment modality (Lin and Baehrecke, 2015). Dysfunctional autophagy contributes too many diseases. Large-scale genomic analysis of human cancers indicates that the loss or mutation of core autophagy genes (Atg) is uncommon, whereas oncogenic events that activate autophagy and lysosomal biogenesis have been identified. Thus, the role of autophagy in cancer is determined by nutrient availability, microenvironment stress, and the presence of an immune system (Amaravadi *et al.*, 2016).

Defective autophagy is implicated in tumorigenesis, as the essential autophagy regulator *Beclin 1* is monoallelically deleted in human breast, ovarian and prostate cancers, and *Beclin 1^{+/*}* mice are tumor-prone. Cellautonomous mechanisms, involving protection of genome integrity and stability, and a non-cell-autonomous mechanism, involving suppression of necrosis and inflammation, have been discovered so far. Autophagy inhibition concurrently with chemotherapy or radiotherapy has emerged as a novel approach in cancer treatment, as autophagy-competent tumor cells depend on autophagy for survival under drug- and radiationinduced stress (Yin *et al.*, 2016; Mizushima and Klionsky 2007; and Baehrecke, 2005). Lower Beclin 1 protein expression, as compared to Beclin 1 levels in normal adjacent breast tissue, was confirmed in a small series of human breast tumors but any correlation between allelic *Beclin 1* loss, and thus defective autophagy, and clinical outcome in breast cancer remains to be investigated (Liang *et al.*, 1999; Feng *et al.*, 2007; and Maiuri *et al.*, 2007).

The mechanism by which autophagy defects lead to accelerated tumorigenesis is not readily apparent, especially given the well-documented pro-survival function of autophagy, which prolongs both normal and tumor cell survival under metabolic stress (Mariño *et al.*, 2007). The protein kinases that are known to target this transcriptional activation domain of p53 can be roughly divided into two groups. A first group of protein kinases belongs to the MAPK family (JNK1-3, ERK1-2, p38 MAPK), which is known to respond to several types of stress, such as membrane damage, oxidative stress, osmotic shock, heat shock, etc. A second group of protein kinases (ATR, ATM, CHK1 and CHK2, DNA-PK, CAK, TP53RK) is implicated in the genome growth (Lumb and Sansom, 2015).

Regulators of apoptosis, such as anti-apoptosis protein Bcl-2/Bcl-xL and the pro-apoptosis protein family, BH3-only proteins, interact with Beclin 1 and can modulate autophagy. The anti-apoptotic protein Bcl-2 binds to Beclin 1 under non-stress conditions and inhibits autophagy in the ER, whereas the BH3-only protein Bad, BNIP3 and BH3 mimetics, such as ABT737 competitively inhibit the interaction between Beclin 1 and Bcl-2/Bcl-xL and stimulate autophagy. Constitutive activation of the PI3K/AKT/mTOR axis is a prototypic survival mechanism commonly encountered in human cancer (Lo Piccolo *et al.*, 2008).

Diverse cellular events, such as loss of the tumor suppressors phosphatase and tensin-homolog deleted on chromosome 10 (PTEN) and tuberous sclerosis complex (TSC) 1 and TSC2, amplification or mutation of class I PI3K, overexpression of AKT, constitutive activation of tyrosine kinase growth factor receptors and exposure to carcinogens, can all result in abnormal activation of this pathway and, ultimately, in autophagy suppression (Arico *et al.*, 2013; and Ueno *et al.*, 2008), (Figure 4).

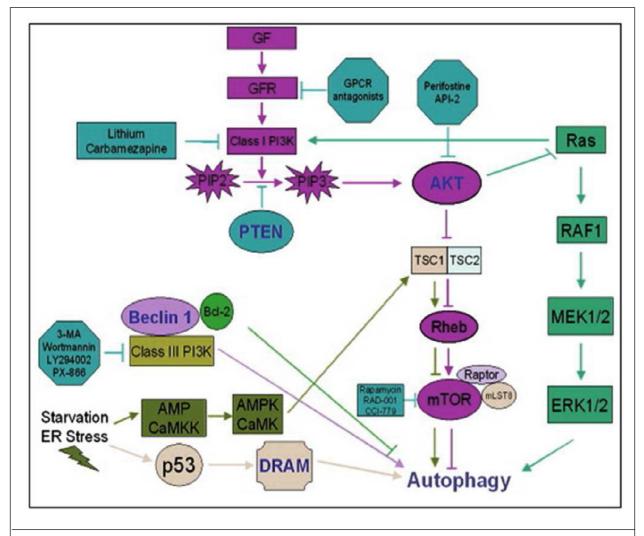


Figure 4: A first group of protein kinases belongs to the MAPK family (JNK1-3, ERK1-2, p38 MAPK), which is known to respond to several types of stress, such as membrane damage, oxidative stress, osmotic shock, heat shock, (Chen V, Karantza-Wadsworth V. Role and regulation of autophagy in cancer. Biochim. Biophys. Acta. 2009; 1793(9): 1516-23)

Two pathways are responsible for autophagy activation in response to starvation and ER stress: (a) mediated by AMPK and Ca-MKK; and (b) involving p53 and damage-regulated autophagy modulator (DRAM) activation. In this context, Ras exhibits an autophagy inhibitor (via class I PI3K activation) and an autophagy activator (via the RAF1/MEK1/2/ERK1/2 pathway) (Chen and Karantza-Wadsworth, 2009). Several mutations or loss of PTEN function affect the activity of lipid phosphatase, which leads to the development of a variety of cancers. Of the three residues in the PTEN component, R-335 was found to be most important to interact with cellular membrane, in common with several other germline mutations, and was associated with inherited cancer (Lumb and Sansom, 2015), (Figure 5).

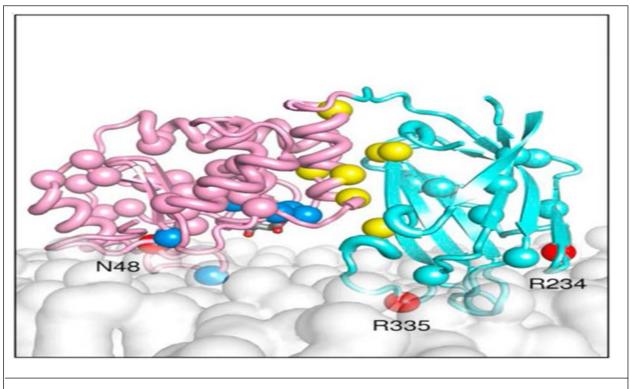


Figure 5: Fosfatase tensin-homolog (PTEN), in membrane location interacting in clinically important mutations (Lumb CN, Sansom MSP. HIF-1á pathway: role, regulation and intervention for cancer therapy. Defining the Membrane-Associated State of the PTEN Tumor Suppressor Protein. Acta Pharm Sin B. 2015 Sep; 5(5): 378-89).

6. Autophagy in Immunological Tolerance

Many studies demonstrated autophagy's contribution to the presentation of cytosolic antigens in association with MHC class II molecules, playing an important role not only in the acquired immune response but also in the maintenance of self-tolerance Mechanisms of central (in the primary lymphoid organs) and peripheral tolerance (in peripheral tissues) physiologically prevent immune responses to self-antigens (Starr *et al.*, 2003). During T cells development in the thymus, the recognition of peptide–MHC molecules on the surface of thymic epithelial cells (TECs) ensures that only thymocytes restricted to MHC molecules, and specific for non-self (foreign) antigens, will survive and continue their maturation. Emerging evidence indicates that autophagy contributes to the maintenance of the central tolerance mechanism (Lleo *et al.*, 2007).

Was recently revealed that there had been an alteration in the selection of the T cell receptor (TCR) restricted to MHC class II in mice transplanted with Atg5–/– thymus. Autophagy defects, in association with a consequent loss of self-tolerance, could be the reason of multiple signs of autoimmunity reported in these animals (Mizushima *et al.*, 2004; and Nedjic *et al.*, 2005). These two opposite results probably depend on the different approach used to inhibit autophagy in the thymus, thus further investigations are necessary (Sukseree *et al.*, 2012). The involvement of autophagy in the presentation of self-antigens to immature T cells in the thymus was first analyzed by Kasai and colleagues, who showed a colocalization of LC3-II with the lysosomal compartment in which MHC–peptide complexes are formed. More recently, Aichinger *et al.* (2013) demonstrated that autophagy is essential for endogenous antigen-loading onto MHC class II of TECs for negative selection (Aichinger *et al.*, 2013).

7. Autophagy in Lymphocytes Homeostasis

Peripheral immune cells play an important role in the perpetuation of autoimmunity by sustaining systemic inflammation status and by participating in the extension of joint destruction mechanisms. Many studies demonstrated that autophagy allows T and B lymphocytes to survive in conditions of nutrients deprivation or during stress stimuli (Gianchecchi *et al.*, 2014). Mice lacking Atg5 do not survive and have a reduction of peripheral T cells, showing how autophagy is essential for their survival. Since cytoplasmic calcium levels are

essential for TCR-signaling pathways activation, autophagy-dependent calcium flux regulation could influence T lymphocytes activation. It has been demonstrated that CD4+ and CD8+ Atg5"/" cells are not able to properly proliferate following TCR stimulation (Pua *et al.*, 2014). Moreover, the inhibition of autophagy causes defects in T cell activation. In fact, deletion of Atg7 results in decreased in IL-2 mRNA level and ATP generation, suggesting that autophagy is required to ensure appropriate energy level for T cell activation (Hubbard *et al.*, 2010). Similar data were obtained also on B lymphocytes, demonstrating that autophagy is essential for the maturation process and for the subsequent maintenance of B lymphocytes repertoire in the periphery (Miller *et al.*, 2008; and Arsov *et al.*, 2011).

8. Autophagy Inhibitors and Therapeutic Implications (Therapeutic Strategy)

According to multiple clinical trials, depending on the type of tumor and its stage of development, activation or inactivation of autophagy may contribute differently to tumorigenesis (Marinkoviæ *et al.*, 2018). In this regard, if increased autophagy confers tumor resistance to death-inducing agents, its inhibition will allow an improved response to treatment (Carew *et al.*, 2012). There are two types of autophagy inhibitors: the early stage which blocks the formation of auto phagosomes (3-methyladenine-3 MA, wortmannin and LY294002) and late stage inhibitors present in the auto phagosome lysosome fusion and degradation phases (chloroquine-CQ, hydroxychloroquine- HCQ (Grácio *et al.*, 2017).

The study of preclinical models for the inhibition of pro-survival autophagy by genetic or pharmacological means has highlighted the possibility of tumor cell destruction and the onset of death of apoptotic cells, (Marino *et al.*, 2014), but also the fact that stress-induced autophagy in tumor cells can lead to resistance to treatment and tumor latency with eventual regeneration and tumor progression (Lu *et al.*, 2008). Several results from studies indicate that an optimal combination of inductors or inhibitors of autophagy (CO, HCO) with chemo or radiotherapy can be successful approaches, being already approved by the Food Dug Administration (FDA) (Solitro and MacKeigan 2016; Boone *et al.*, 2015; and Pan *et al.*, 2016).

Based on the fact that approximately 70% of clinical trials focus on the role of autophagy in cancer, it indicates that the potential for autophagy modulation in cancer treatment is promising. Clinical studies involving autophagy modulation in cancers have been designed to evaluate the effect of autophagy inhibition in combination with other conventional therapies. Only a small portion of the clinical studies on lung cancer, glioblastoma, pancreas, melanoma, breast and prostate cancer test CQ / HCQ as monotherapy. Current clinical studies (pancreatic adenocarcinoma) have shown that autophagy inhibition by HCQ as monotherapy is not sufficient (Wolpin *et al.*, 2014). Thus, the use of autophagy inhibitors in combination with chemotherapy can suppress tumor growth and trigger cell death in a higher percentage than both chemotherapy alone in vitro and in vivo Table 1, (Yang *et al.*, 2011; and Amaravadi *et al.*, 2011).

Table 1: Autophagy modulators apply in preclinical and clinical studies/ trials for cancer therapy. (Amaravadi RK, Lippincott-Schwartz J, Yin XM, Weiss WA, Takebe T. Principles and Current Strategies for Targeting Autophagy for Cancer Treatment. Clin Cancer Res. 2011; 17(4): 654–66).		
Cancer type	Autophagy modulators	Model tested/clinical trial phase
Pancreas cancer		
Stage IIb or III pancreatic adenocarcinoma	HCQ+gemcitabine	Phase I/II
Advanced metastatic pancreatic adenocarcinoma	HCQ+gemcitabine/abraxane	Phase I/II
Metastatic pancreatic adenocarcinoma	HCQ; Gemcitabine, nab-paclitaxel	Phase II
Hepatocellular cancers		
HCC after liver transplantation	Sirolimus; RAD001; Sorafenib+HCQ	Phase II; Phase III; Phase II
Advanced HCC	Sirolimus; Sorafenib	Phase II/III,

The American Brain Tumor Consortium initiated a phase I / II trial for glioblastoma patients using HCQ, temozolomide and radiation highlighted the level of inhibition of HCQ dependent autophagy using an electron microscopy test on blood serum mononuclear cells (Rosenfeld *et al.*, 2010). Electronic micrographs of peripheral blood mononuclear cells from a glioma patient included in the Phase I/II study using temozolomide, radiation and hydroxychloroquine TO demonstrates the antitumor activity of this combination.

Induction of autophagy is another way that can help improve the effect of anticancer therapies when autophagy is cytotoxic by inducing cell death – apoptosis. They have been described in the literature and use a range of natural drugs/extracts to induce cell death mediated by autophagy in various cancer cells (Bristol *et al.*, 2012; Selvaraj *et al.*, 2016; Acharya *et al.*, 2011; Turcotte *et al.*, 2018; and Lima *et al.*, 2016).

Some recent studies describe the important role of autophagy in regulating immune recognition and responding to tumor cell immunogenicity, involvement in tumor antigen, processing and subsequent activation of effector T cells (Viry *et al.*, 2014). In this direction, the adjuvant potential in stimulating the antitumoral immune response was experimentally demonstrated in mice with pulmonary carcinoma and melanoma (Li *et al.*, 2001).

9. Conclusion

Autophagy defects are associated with susceptibility to metabolic stress, DNA damage, accumulation, genomic instability, and accelerated tumorigenicity, and these observations lead to the predictions that autophagy stimulation may preserve cellular fitness and genome integrity, and thus prevent cancer, and that tumors with chronic autophagy deficiency may be particularly sensitive to certain anticancer agents, such as DNA-damaging and anti-angiogenic drugs. Inhibition of autophagy concurrently with treatment may augment the antitumor activity, and thus the efficacy, of radiation and/or anticancer drugs. Ultimately, pharmacologic manipulation of autophagy for cancer prevention and treatment will depend on our ability to successfully recognize the functional status of autophagy in tumors and on the availability of specific autophagy modulators.

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References

- Acharya, B.R., Bhattacharyya, S., Choudhury, D. and Chakrabarti, G (2011). The microtubule depolymerizing agent naphthazarin induces both apoptosis and autophagy in A549 lung cancer cells. *Apoptosis*. 16, 930-939.
- Aichinger, M., Wu, C., Nedjic, J. and Klein, L. (2013). Macroautophagy substrates are loaded onto MHC class II of medullary thymic epithelial cells for central tolerance. *J. Exp. Med.* 210 (2), 287-300.
- Alessandri, C., Barbati, C., Vacirca, D., Piscopo, P., Confaloni, A., Sanchez, M., *et al.* (2012). T lymphocytes from patients with systemic lupus erythematosus are resistant to induction of autophagy. *FASEB J.* 26 (11), 4722-4732.
- Amaravadi, K.R., Yu, D., Lum, J.J., Bui, T., Christophorou, A.M. and Evan, I.G., *et al.* (2007). Autophagy inhibition enhances therapy-induced apoptosis in a Myc-induced model of lymphoma. *J. Clin. Invest.* 117, 326-336.
- Amaravadi, R., Kimmelman, A.C. and White, E. (2016). Recent insights into the function of autophagy in cancer. *Genes Dev.* 30 (17), 1913-1930.
- Amaravadi, R.K., Lippincott-Schwartz, J., Yin, X.M., Weiss, W.A. and Takebe, T. (2011). Principles and Current Strategies for Targeting Autophagy for Cancer Treatment. *Clin Cancer Res.* 17, (4), 654-666.
- Arico, S., Petiot, A., Bauvy, C., Dubbelhuis, F.P., Meijer, J.A. and Codogno, P. (2013). The tumor suppressor PTEN positively regulates macroautophagy by inhibiting the phosphatidylinositol 3-kinase/protein kinase B pathway. J. Biol. Chem. 276, 5243-5246.
- Arsov, I., Adebayo, A., Kucerova-Levisohn, M., Haye, J., MacNeil, M. and Papavasiliou, F.N., *et al.* (2011). A role for autophagic protein beclin1 early in lymphocyte development. *J. Immunol.* 186 (4), 2201-2229.

Baehrecke, E.H. (2005). Autophagy: Dual roles in life and death?. Nat. Rev. Mol. Cell. Biol. 6 (6), 505-510.

- Barbati, C., Alessandri, C., Vomero, M., Vona, R., Colasanti, T., Vacirca, *et al.* (2015). Autoantibodies specific to D4GDI modulate Rho GTPase mediated cytoskeleton remodeling and induce autophagy in T lymphocytes. *J. Autoimmun.* 58, 78-89.
- Boone, B.A., Bahary, N., Zureikat, A., Moser, A.J. and Normolle, D.P. *et al.* (2015). Safety and biologic response of pre-operative autophagy inhibition in combination with gemcitabine in patients with pancreatic adenocarcinoma. *Ann. Surg. Oncol.* 22, 4402-4410.
- Boya, P., Gonzalez-Polo, A.R., Casares, N., Perfettini, L.J., Dessen, P.N. and Larochette, D. (2005). Inhibition of macroautophagy triggers apoptosis. *Mol. Cell. Biol.* 25, 1025-1040.
- Bristol, M.L., Di, X., Beckman, M.J., Wilson, E.N. and Henderson, S.C., *et al.* (2012). Dual functions of autophagy in the response of breast tumor cells to radiation: cytoprotective autophagy with radiation alone and cytotoxic autophagy in radiosensitization by vitamin D 3. *Autophagy*. 8, 739-753.
- Carew, J.S., Kelly, K.R. and Nawrocki S.T. (2012). Autophagy as a target for cancer therapy: new developments. *Cancer Manag. Res.* (90), 40: 357-365.
- Chen, N. and Karantza-Wadsworth, V. (2009). Role and regulation of autophagy in cancer. *Biochim. Biophys. Acta*. 1793 (9), 1516-1523.
- Colasanti, T., Vomero, M., Alessandri, C., Barbati, C., Maselli, A., Camperio, C., *et al.* (2014). Role of alphasynuclein in autophagy modulation of primary human T lymphocytes. *Cell. Death Dis.* 5: e1265. Doi:10.1038/cddis.2014.211.
- Di Bartolomeo, S., Corazzari, M., Nazio, E., Oliverio, S., Lisi, G. and Antonioli, M. *et al.* (2013). The dynamic interaction of AMBRA1 with the dynein motor complex regulates mammalian autophagy. *J. Cell. Biol.* 191 (1), 155-168.
- Feng, W., Huang, S., Wu, H. and Zhang, M. (2007). Molecular basis of Bcl-xL's target recognition versatility revealed by the structure of Bcl-xL in complex with the BH3 domain of Beclin-1. *J. Mol. Biol.* 372, 223-235.
- Feng, Z., Hu, W., Stanchina, E., Teresky, A.K., Jin, S. and Lowe, S., et al. (2007). The regulation of AMPK beta1, TSC2, and PTEN expression by p53: stress, cell and tissue specificity, and the role of these gene products in modulating the IGF-1-AKT-mTOR pathways. *Cancer. Res.* 67, 3043-3053.
- Feng, Z., Zhang, H., Levine, J.A. and Jin, S. (2015). The coordinate regulation of the p53 and mTOR pathways in cells. *Proc. Natl. Acad. Sci. USA.* 102, 8204-8209.
- Fimia, G.M., Stoykova, A., Romagnoli, A., Giunta, L., Di Bartolomeo, S., Nardacci, R., *et al.*, (2007). Ambra1 regulates autophagy and development of the nervous system. *Nature*. 447 (7148), 1121–1125.
- Fougeray, S. and Pallet, N. (2015). Mechanisms and biological functions of autophagy in diseased and ageing kidneys. *Nat. Rev. Nephrol.*, 11 (1), 34-45. doi: 10.1038/nrneph.2014.201.
- Galluzzi, L., Pietrocola, F., Bravo-San Pedro, J.M., Amaravadi, R.K., Baehrecke, E.H., Cecconi, F., *et al.* (2015). Autophagy in malignant transformation and cancer progression. *Embo J.* 34 (7), 856-880.
- Gianchecchi, E., Delfino, D.V., Fierabracci, A. (2014). Recent insights on the putative role of autophagy in autoimmune diseases. *Autoimmun. Rev.* 13 (3), 231-241.
- Grácio, D., Magro, F., Lima, R.T. and Máximo V. (2017). An overview on the role of autophagy in cancer therapy. *Hematol. Med. Oncol.* 2, 1: 1-4. doi: 10.15761/HMO.1000117
- Hosokawa, N., Hara, T., Kaizuka, T., Kishi, C., Takamura, A., Miura, Y., *et al.* (2009). Nutrient-dependent mTORC1 association with the ULK1-Atg13- FIP200 complex required for autophagy. *Mol. Biol. Cell.* 20 (7), 1981-1991.
- Hubbard, V.M., Valdor, R., Patel, B., Singh, R., Cuervo, A.M. and Macian, F. (2010). Macroautophagy regulates energy metabolism during effector T cell activation. *J. Immunol.* 185 (12), 7349-7357.
- Jin, S., DiPaola, S.R., Mathew, R. and White, E. (2007). Metabolic catastrophe as a means to cancer cell death. J. Cell. Sci. 120, 379-383.

- Klionsky, D.J., Abdelmohsen, K., Abe, A., Abedin, M.J., Abeliovich, H., Acevedo Arozena, A., *et al.* (2016). Guidelines for the use and interpretation of assays for monitoring autophagy (3rd Edition). *Autophagy*. 12 (1), 1-222.
- Levine, B., Sinha, S. and Kroemer, G. (2008). Bcl-2 family members: dual regulators of apoptosis and autophagy. *Autophagy*. 4 (5), 600-606.
- Levy, J.M., Thompson, J.C., Griesinger, A.M., Amani, V., Donson, A.M., Birks, D.K., et al. (2014). Autophagy inhibition improves chemosensitivity in BRAF (V600E) brain tumors. *Cancer Discov.* 4 (7), 773-780.
- Li, Y., Wang, L.X., Pang, P., Cui, Z., Aung, S., *et al.* (2011). Tumor-derived autophagosome vaccine: mechanism of cross-presentation and therapeutic efficacy. *Clin. Cancer Res.* 17, 7047-7057.
- Liang, H.X., Jackson, S., Seaman, M., Brown, K., Kempkes, B. and Hibshoosh, H. (1999). Induction of autophagy and inhibition of tumorigenesis by beclin 1. *Nature*. 402, 672-676.
- Lima, R. T., Sousa, D., Paiva, A. M., Palmeira, A. and Barbosa, J., *et al.* (2016). Modulation of autophagy by a thioxanthone decreases the viability of melanoma. *Molecules.* 21 (10), 1343-1345.
- Lin, L. and Baehrecke. E.H. (2015). Autophagy, cell death, and cancer. *Mol. Cell. Oncol.* 2 (3), e985913. doi: 10.4161/23723556.2014.985913
- Lleo, A., Invernizzi, P., Selmi, C., Coppel, R.L., Alpini, G. and Podda, M., et al. (2007). Autophagy: highlighting a novel player in the autoimmunity scenario. J. Autoimmun. 29 (2-3), 61-68.
- Lo Piccolo, J., Blumenthal, M.G., Bernstein. B.W. and Dennis, A.P. (2008). Targeting the PI3K/Akt/mTOR pathway: Effective combinations and clinical considerations. Drug Resist. Updat. 11, 32-50.
- Lu, Z., Luo, R.Z., Lu, Y., Zhang, X., Yu, Q. and Khare, S., et al. (2008). The tumor suppressor gene ARHI regulates autophagy and tumor dormancy in human ovarian cancer cells, J. Clin. Invest. 118, 3917–3929.
- Lumb, C.N. and Sansom, M.S.P. (2013). Defining the membrane-associated state of the PTEN tum or suppressor protein. *Biophys J.* 104 (3), 613-621.
- Lumb, C.N. and Sansom, M.S.P. (2015). HIF-1α pathway: Role, regulation and intervention for cancer therapy. Defining the Membrane-Associated State of the PTEN Tumor Suppressor Protein. *Acta Pharm Sin B*. 5 (5), 378-389.
- Madeo, F., Zimmermann, A., Maiuri, M.C. and Kroemer, G. (2015). Essential role for autophagy in life span extension. *J. Clin. Invest.* 125 (1), 85-93.
- Maiuri, C.M., Criollo, A., Tasdemir, E., Vicencio, M.J., Tajeddine, N.J. and Hickman, A. (2007). BH3-only proteins and BH3 mimetics induce autophagy by competitively disrupting the interaction between Beclin 1 and Bcl-2/Bcl-X(L). *Autophagy*. 3, 374-376.
- Marinkoviæ, M., Šprung, M., Buljubašiæ, M. and Novak, I. (2018). Autophagy modulation in cancer: Current knowledge on action and therapy. *Oxid. Med Cell. Longev.* 5, 1-18. https://doi.org/10.1155/2018/ 8023821
- Marino, G., Niso-Santano, M., Baehrecke, H.E. and Kroemer, G. (2014). Self-consumption: the interplay of autophagy and apoptosis. *Nature Reviews Molecular Cell Biology*. 15 (2), 81-94.
- Mariño, G., Salvador-Montoliu, N., Fueyo, A., Knecht, E., Mizushima, N., López-Otín. C. (2007). Tissue-specific autophagy alterations and increased tumorigenesis in mice deficient in Atg4C/autophagin-3. *J. Biol. Chem.* 282, 8573-8583.
- Masoud, G.N. and Wei Li W. (2015). HIF-1α pathway: role, regulation and intervention for cancer therapy. *Acta Pharm Sin B.* 5 (5), 378-389.
- Miller, B.C., Zhao, Z., Stephenson, L.M., Cadwell, K., Pua, H.H. and Lee, H.K., *et al.* (2008). The autophagy gene Atg5 plays an essential role in B lymphocyte development. *Autophagy*. 4 (3), 309-314.
- Mizushima, N. and Klionsky, D. (2007). Protein turnover via autophagy: implications for metabolism. *Annu. Rev.* 27, 19-40.
- Mizushima, N., Yamamoto, A., Matsui, M., Yoshimori, T. and Ohsumi Y. (2004). *In vivo* analysis of autophagy in response to nutrient starvation using transgenic mice expressing a fluorescent autophagosome marker. *Mol. Biol. Cell.* 15 (3), 1101–1111.

- Muller, S., Brun, S., René, F., de Sèze, J., Loeffler, J.P. and Jeltsch-David, H. (2017). Autophagy in neuroinflammatory diseases. *Autoimmun Rev.*, 16 (8), 856-874.
- Nedjic, J., Aichinger, M., Emmerich, J., Mizushima, N. and Klein, L. (2008). Autophagy in thymic epithelium shapes the T-cell repertoire and is essential for tolerance. *Nature*, 455 (7211), 396-400.
- O'Farrell, F., Rusten, T.E. and Stenmark. H. (2013). Phosphoinositide 3-kinases as accelerators and brakes of autophagy, *FEBS J.*, 280 (24), 6322-6337.
- Ohsumi, Y. (2014). Historical landmarks of autophagy research. Cell. Res. 24 (1), 9-23.
- Pan, H., Chen, L. and Xu, Y., et al. (2016). Autophagy-associated immune responses and cancer immunotherapy. Oncotarget. 7 (16), 21235-21246.
- Pua, H.H., Dzhagalov, I., Chuck, M., Mizushima, N. and He, Y.W. (2014). A critical role for the autophagy gene Atg5 in T cell survival and proliferation. *J. Exp. Med.* (1), 25-31.
- Ravikumar, B., Moreau, K., Jahreiss, L., Puri, C. and Rubinsztein, D.C. (2010). Plasma membrane contributes to the formation of pre-autophagosomal structures. *Nat. Cell. Biol.*, 12 (8), 747-757.
- Rosenfeld, M.R., Brem, S., Mikkelsen, T., Wang, D., Piao, S., Davis, L.E., O'Dwyer, P.J. and Amaravadi, R.K. (2010). Pharmacokinetic analysis and pharmacodynamic evidence of autophagy inhibition in patients with newly diagnosed glioblastoma treated on a phase I trial of hydroxychloroquine in combination with adjuvant temozolomide and radiation (ABTC 0603). J. Clin. Oncol. 28 (15_suppl), 3086-3086.
- Selvaraj, S., Sun, Y., Sukumaran, P. and Singh, B.B. (2016). Resveratrol activates autophagic cell death in prostate cancer cells via downregulation of STIM1 and the mTOR pathway. *Mol. Carcinog.* 55, 818-830.
- Solitro, A.R. and MacKeigan, J.P. (2016). Leaving the lysosome behind: novel developments in autophagy inhibition. *Future Med. Chem.* 8, 73-86.
- Starr, T.K., Jameson, S.C., Hogquist, K.A. (2003). Positive and negative selection of T cells. *Annu. Rev. Immunol.* 21, 139-176.
- Sukseree, S., Mildner, M., Rossiter, H., Pammer, J., Zhang, C.F. and Watanapokasin, R., *et al.* (2012). Autophagy in the thymic epithelium is dispensable for the development of self-tolerance in a novel mouse model. *PLoS One.* 7 (6), e38933. doi: 10.1371/journal.pone.0038933
- Takahashi, Y., Meyerkord, C.L., Hori, T., Runkle, K., Fox, T.E., Kester, M. *et al.* (2011). Bif-1 regulates Atg9 trafficking by mediating the fission of Golgi membranes during autophagy. *Autophagy*. 7 (1), 61–73.
- Tang, J., Di, J., Cao, H., Bai, J., Zheng, J. (2015). p53-mediated autophagic regulation: A prospective strategy for cancer therapy. *Cancer Lett.* 363 (2), 101-107.
- Tasdemir, E., Maiuri, C.M., Galluzzi, L., Vitale, J. and Djavaheri-Mergny, M., *et al.* (2008). Regulation of autophagy by cytoplasmic p53. *Nat. Cell. Biol.*, 10, 676-687.
- Tracy, K., Dibling, C.B., Spike, T.B., Knabb, J.R., Schumacker, P. and Macleod K.F. (2007). BNIP3 is an RB/E2F target gene required for hypoxia-induced autophagy. *Mol. Cell. Biol.* 27, 6229-6242.
- Turcotte, S., Chan, D.A., Sutphin, P.D., Hay, M.P. and Denny, W.A., et al. (2018). A molecule targeting VHLdeficient renal cell carcinoma that induces autophagy. *Cancer Cell*. 14, 90-102.
- Udristioiu, A. and Nica-Badea, D. (2018). Signification of protein p-53 isoforms and immune therapeutic success in chronic lymphocytic leukemia. *Biomedicine & Pharmacotherapy*. 106, 50-53.
- Ueno, T., Sato, W., Horie, Y., Komatsu, M., Tanida, I., and Yoshida, M., *et al.* (2008). Loss of Pten, a tumor suppressor, causes the strong inhibition of autophagy without affecting LC3 lipidation. *Autophagy*. 4, 692-699.
- Viry, E., Paggetti, J., Baginska, J., Mgrditchian T., Berchem G., et al. (2014). Autophagy: An adaptive metabolic response to stress shaping the antitumor immunity. *Biochem. Pharmacol.* 92, 31-42.
- Vomero, M., Barbati, C., Colasanti, T., Perricone, C., Novelli, L., Ceccarelli, F. *et al.* (2018). Autophagy and rheumatoid arthritis: Current knowledges and future perspectives. *Front Immunol.*, 9, 1577. doi: 10.3389/ fimmu.2018.01577.
- Wolpin, B. M., Rubinson, D. A. and Wang, X., *et al.* (2014). Phase II and pharmacodynamic study of autophagy inhibition using hydroxychloroquine in patients with metastatic pancreatic adenocarcinoma,. *The Oncologist.* 19 (6), 637-638.

- Yang, Z. J., Chee, C. E., Huang, S. and Sinicrope, F. A. (2011). The role of autophagy in cancer: Therapeutic implications. *Mol. Cancer. Ther.* 10 (9), 1533-1541. doi: 10.1158/1535-7163.MCT-11-0047.
- Yin, Z., Pascual, C. and Klionsky, D.J. (2016). Autophagy: machinery and regulation. *Microb. Cell.* 3 (12), 588-596.
- Yla-Anttila, P., Vihinen, H., Jokitalo, E. and Eskelinen, E-L. (2009). 3D tomography reveals connections between the phagophore and endoplasmic reticulum. *Autophagy*. 5 (8), 1180-1185.
- Yu, L., Chen, Y. and Tooze, S.A. (2018). Autophagy pathway: Cellular and molecular mechanisms. *Autophagy*. 14 (2), 207-215. doi: 10.1080/15548627.2017.137883
- Yuan, T.L. and Cantley, L.C. (2008). PI3K pathway alterations in cancer: Variations on a theme. *Oncogene*. 27, 5497-5510.

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